Removal and Rebound Kinetics of Cystatin C in High-Flux Hemodialysis and Hemodiafiltration

Enric Vilar,*† Capella Boltiador,*§ Adie Viljoen,*§ Ashwini Machado,* and Ken Farrington*†

Abstract
Background and objectives Cystatin C is a 13.3 kD middle molecule of similar size to β2-microglobulin and a marker of GFR in CKD. This study aimed to determine cystatin C kinetics in hemodialysis to understand whether blood concentrations may predict residual renal function and middle-molecule clearance.

Design, setting, participants, & measurements Cystatin C removal and rebound kinetics were studied in 24 patients on high-flux hemodialysis or hemodiafiltration. To determine whether cystatin C concentrations are predictable, an iterative two-pool mathematical model was applied.

Results Cystatin C was cleared effectively, although less than β2-microglobulin (reduction ratios ± SD are 39% ± 11 and 51% ± 11). Cystatin C rebounded to 95% ± 5% of predialysis concentration by 12 hours postsedialysis. The two-pool kinetic model showed excellent goodness of fit. Modeled extracellular cystatin C pool volume is smaller than that predicted, comprising 25.5% ± 9.2 of total body water. Iterated parameters, including nonrenal clearance, showed wide interindividual variation. Modeled nonrenal clearance was substantially higher than renal clearance in this population at 25.1 ± 6.6 ml/min per 1.73 m² body surface area.

Conclusions Plasma cystatin C levels may be used to measure middle-molecule clearance. Levels rebound substantially postdialysis and plateau in the interdialytic period. At low GFR, nonrenal clearance predominates over renal clearance, and its interindividual variation will limit use of cystatin C to predict residual renal function in advanced kidney disease.


Introduction
Cystatin C is a lysosomal protease and cysteine proteinase inhibitor produced by all nucleated cells (1,2). It is freely filtered by glomeruli and metabolized by proximal tubules (3,4). Considerable interest has developed in use of cystatin C as a marker of GFR (5). Cystatin C-based GFR estimates do not require correction for muscle bulk, sex, and race (6–8). Cystatin C predicts mortality in CKD independent of GFR (9). There has been interest in use of cystatin C as a marker of residual renal function (RRF) in both acute renal failure and ESRD (10–12). Previous data suggest that, at low GFR, the close relationship of cystatin C with GFR weakens (11). However, if cystatin C kinetics are predictable, then it may be possible to use parameters, such as predialysis cystatin C, to predict RRF. This use would have wide-ranging benefits and allow estimation of RRF without the need for interdialytic urine collection (13).

There is increasing interest in techniques that maximize middle-molecule clearance, such as hemodiafiltration (HDF) (14,15). Clearance of these molecules may be an important marker of adequate treatment in both peritoneal dialysis (10,16) and hemodialysis (HD) (11,17). Cystatin C has been proposed as a candidate for such a marker (18–20), but there are little data describing its removal and rebound kinetics in the maintenance HD population.

We aimed to determine plasma cystatin C kinetics in high-flux HD and HDF during dialysis and the interdialytic period to establish the relationship of cystatin C levels to dialyzer and renal clearance. Cystatin C generation rate (G) and nonrenal clearance (KNR) have been previously estimated (21). Because the sieving coefficient of cystatin C is close to unity (4), the renal clearance (Kd) approximates GFR. Intercompartmental transfer rates are unknown, but limited information suggests that the major pool is extracellular.

To determine cystatin C kinetics in high-flux HD and HDF and whether plasma concentrations are predictable, we developed a two-pool mass transfer iterative computer model. This model used previously available data on G, KNR, measured GFR, and plasma cystatin C levels sampled throughout the HD cycle.

Materials and Methods
Study Design and Setting
After approval by a local research ethics committee and in accordance with the Declaration of Helsinki, 24 patients on either chronic high-flux HD or HDF at the Lister Hospital in Stevenage, United Kingdom gave
HD and HDF Procedures

Patients were dialyzed using an incremental HD algorithm, where the target total Kt/V was 1.2 (22). GFR was estimated as the mean of urea and creatinine clearance calculated from a midweek 48-hour interdialytic urine collection performed during the study period ±7 days. Patients with a urine output < 100 ml/d were classified as having 0 GFR. Urea and creatinine clearance were calculated using the formula

\[
\text{Clearance} = \frac{2(U_{ID} + V_{ID})}{\text{I}_{ID}(C_{\text{post}} + C_{\text{pre}})}
\]

where \(U_{ID}\) is the urinary concentration of the relevant solute (millimoles per liter or micromoles per liter), \(V_{ID}\) is the urine collection volume (milliliters), \(I_{ID}\) is the duration of interdialytic period (minutes), and \(C_{\text{post}}\) and \(C_{\text{pre}}\) are the post- and predialysis plasma concentrations of the relevant solute (millimoles per liter or micromoles per liter).

Dialysis data recorded included blood flow (\(Q_b\)), dialysis fluid flow (\(Q_d\)), ultrafiltration volume (UF), and body size parameters.

Sampling of Plasma for Clearance Molecules

Samples were obtained during dialysis from the arterial needle immediately before the first dialysis session (baseline) and at four intervals equal to 20% of the dialysis time. The end of dialysis sample was taken 30 seconds after slowing the blood pump to 50 ml/min (23). Postdialysis samples were obtained at 1, 2, 4, 8, 12, and 24 hours and immediately before the subsequent dialysis session. For the first five (pilot) patients, additional postdialysis samples were taken at 2, 15, and 30 minutes (the washback was after the 2-minute sample).

All samples were analyzed for urea, creatinine, \(\beta_2\)-microglobulin, and cystatin C as described in Supplemental Material.

Design of Two-Pool Mass Transfer Model of Cystatin C Kinetics

Model Design. Model design is shown in Figure 1. Cystatin C was assumed to be distributed in intracellular and extracellular water with concentrations \(C_i\) and \(C_e\), respectively. Mass transfer rate between pools was assumed to equal the mass transfer coefficient for the intracellular/extracellular barrier (\(K_{int}\)) multiplied by the difference between \(C_i\) and \(C_e\). \(G\) was assumed to be into intracellular space at a constant rate. Cystatin C removal from extracellular space was assumed to occur by three routes: \(K_R\), \(K_{NR}\), and dialyzer clearance (\(K_D\)). Each of these rates was equal to the product of the clearance constant (\(K_{NR}\) or \(K_D\)) and \(C_e\). The model used a variable volume, with ultrafiltration volume being total fluid removed over the dialysis period and assumed to be at a constant rate during HD from the extracellular compartment.

Cystatin C concentration in the intracellular and extracellular pools was calculated from the mass of cystatin C in the respective pool divided by the pool volume (\(V_i\) or \(V_e\)).

The Watson volume (\(V_{\text{Watson}}\)) (24) was assumed to be equal to postdialysis water body (\(TBW_{\text{post}}\)) and calculated using the clinically assessed dry weight. Predialysis total body water (\(TBW_{\text{pre}}\)) was considered to be \(V_{\text{Watson}}\) plus the UF volume (\(TBW_{\text{pre}} = V_{\text{Watson}} + UF\)). \(TBW_{\text{pre}}\) was considered to consist of predialysis extracellular and intracellular water volumes (\(V_{e,\text{pre}}\) and \(V_{i,\text{pre}}\) respectively). A constant (\(ECW_{\text{Fraction}}\)) defined the proportion of \(TBW_{\text{pre}}\) occupied by extracellular water (\(V_{e,\text{pre}}/TBW_{\text{pre}}\)). The fraction of \(TBW_{\text{pre}}\) occupied by intracellular water was, therefore, \(1 - ECW_{\text{Fraction}}\). Equations defining the initial (predialysis) pool volumes were, therefore, \(V_{e,\text{pre}} = ECW_{\text{Fraction}} \times TBW_{\text{pre}}\) and \(V_{i,\text{pre}} = (1 - ECW_{\text{Fraction}}) \times TBW_{\text{pre}}\). Mass transfer in or out of pools in the model was calculated using the integral of the mass transfer rates.

Modeling was performed using Vissim Optimize-Pro v8.0 (Westford, MA), permitting an iterative model to be designed for mass flow between compartments using integration. This environment permits importing of observed data and comparison with modeled plasma cystatin C concentration, and it has previously been used for dialysis kinetic modeling (25).

Iterative Procedure. Certain parameters were taken as constants—either measured or assumed from previously reported data. Unknown parameters were iterated. Constants were cystatin C \(K_D\), \(K_R\), and \(G\). Iterated parameters were \(K_{NR}\), \(ECW_{\text{Fraction}}\), and \(K_{int}\). Published cystatin C clearance data for high-flux dialyzer membranes are limited, but data indicate that it is approximately 10% lower than in vivo \(\beta_2\)-microglobulin clearance (26–28). We, therefore, estimated cystatin C \(K_D\) to be 90% of \(\beta_2\)-microglobulin clearance based on extensively studied \(\beta_2\)-microglobulin clearance data, which took account of modality (HD/HDF), blood flow, and dialyzer size (29). \(K_R\) was assumed
to be equal to measured GFR that was unadjusted for body surface area, which assumed a sieving coefficient of one for cystatin C (4). G was taken as 0.124 mg/min per 1.73 m² body surface area in accordance with previous data (21).

Unknown parameters (KNR, ECWFraction, and Kint) were iterated by optimal fitting of the model to the observed patient data. Estimates of anticipated parameter values were used as initial values. The initial iteration value of KNR was 22.3 ml/min per 1.73 m² body surface area from previous data (21). Simultaneous iteration of both G and KNR was not possible because of their opposite overall effects on the model, which would result in multiple possible solutions. In view of the potential for multiple pool effects for cystatin C, we iterated ECWFraction. The initial iteration value of ECWFraction (ECWFraction_initial_value) was calculated as the predicted predialysis proportion of TBW occupied by extracellular water according to

\[
ECWFraction_{\text{initial value}} = \frac{[UF + (\text{Weight}_{\text{post}} \times \text{factor})]}{[V_{\text{Watson}} + UF]},
\]

where Weight_post is postdialysis weight and factor represents the predicted ratio of extracellular water to body weight, which was taken as 0.239 and 0.214 L/kg in men and women, respectively (30). In one patient with a unilateral below knee amputation, V_Watson was corrected (31), and initial value of ECWFraction was assumed to be 0.4.

The initial value of K_int for each patient was calculated by running a test model for each subject, in which KNR and Kint were assumed to be equal to measured GFR that was unadjusted for body surface area. For each molecule, there was no significant difference in mean concentration between +2- and +4-hours postdialysis time points. Concentrations of urea, creatinine, and β2-microglobulin did not rebound to their pre-dialysis levels during the interdialytic period, whereas for cystatin C, the mean 12-hour postdialysis concentration was 95%±5 of the predialysis concentration, and by 24 hours, it had reached 103%±7 (Figure 2). For patients treated by high-flux HD, the mean cystatin C reduction ratio was

(13 patients on high-flux HD and 11 patients on online HDF). Hospitalized patients were studied before planned discharge after they were considered hemodynamically and clinically stable. Hospitalization had been related to vascular access (21%), infections (25%), cardiovascular problems (38%), and other reasons (16%).

### Clearance of Cystatin C Compared with Small and Middle Molecules

Figures 2 and 3 show observed clearance and rebound kinetics for urea, creatinine, β2-microglobulin, and cystatin C; reduction ratios for each molecule (±SD) were 63%±11, 58%±12, 51%±11, and 39%±11, respectively. For all molecules, a rapid rebound occurred in the first 1 hour after termination of dialysis followed by a shallower rebound. For each molecule, there was no significant difference in mean concentration between +2- and +4-hours postdialysis time points. Concentrations of urea, creatinine, and β2-microglobulin did not rebound to their pre-dialysis levels during the interdialytic period, whereas for cystatin C, the mean 12-hour postdialysis concentration was 95%±5 of the predialysis concentration, and by 24 hours, it had reached 103%±7 (Figure 2). For patients treated by high-flux HD, the mean cystatin C reduction ratio was

### Results

#### Study Population

Patient demographics, comorbidities, and dialysis parameters are detailed in Table 1; 24 patients were recruited (13 patients on high-flux HD and 11 patients on online HDF). Hospitalized patients were studied before planned discharge after they were considered hemodynamically and clinically stable. Hospitalization had been related to vascular access (21%), infections (25%), cardiovascular problems (38%), and other reasons (16%).

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### Table 1. Baseline characteristics, comorbidities, and dialysis parameters in the study population

<table>
<thead>
<tr>
<th></th>
<th>Parameter</th>
</tr>
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<tbody>
<tr>
<td><strong>Demographics</strong></td>
<td></td>
</tr>
<tr>
<td>Age, yr</td>
<td>62.0±14.6</td>
</tr>
<tr>
<td>Men/women, %</td>
<td>46/54</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>67.2±22.1</td>
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<tr>
<td>Comorbidities</td>
<td></td>
</tr>
<tr>
<td>Diabetes mellitus, %</td>
<td>67</td>
</tr>
<tr>
<td>Cardiovascular disease, %</td>
<td>50</td>
</tr>
<tr>
<td>Active malignancy, %</td>
<td>8.3</td>
</tr>
<tr>
<td>C reactive protein, mg/L</td>
<td>22 (62)</td>
</tr>
<tr>
<td><strong>Dialysis parameters</strong></td>
<td></td>
</tr>
<tr>
<td>Dialysis time, min</td>
<td>188±33</td>
</tr>
<tr>
<td>Blood flow (Qb), ml/min</td>
<td>283±43</td>
</tr>
<tr>
<td>Dialysis fluid flow (QD), ml/min</td>
<td>587±139a</td>
</tr>
<tr>
<td>High-flux hemodialysis/ hemodiafiltration, %</td>
<td>54/46</td>
</tr>
<tr>
<td>Ultrafiltration, L</td>
<td>1.49±1.05a</td>
</tr>
<tr>
<td>Estimated in vivo cystatin C</td>
<td>70.3±20.1a</td>
</tr>
<tr>
<td><strong>GFR, ml/min per 1.73 m²</strong></td>
<td>1.43 (0.22–2.33)b</td>
</tr>
<tr>
<td>body surface area</td>
<td></td>
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<tr>
<td>Single pool Kt/V_urea</td>
<td>1.19±0.52</td>
</tr>
<tr>
<td>Total Kt/V_urea (renal+dialyzer)</td>
<td>1.24±0.49</td>
</tr>
<tr>
<td>Vascular access</td>
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<tr>
<td>Arteriovenous fistula, %</td>
<td>12.5</td>
</tr>
<tr>
<td>Vascular catheter, %</td>
<td>87.5</td>
</tr>
</tbody>
</table>

Table 1 represents the sum of residual renal and dialyzer clearance, which was used for incremental dialysis calculations (Materials and Methods).

**a**Mean±SD.

**b**Median (interquartile range).
34%±9%, and for those patients on HDF, it was 44%±11% (P=0.02) (Figure 4). Likewise, for β2-microglobulin, the mean reduction ratio on high-flux HD was 46%±11% compared with 56%±10% on HDF (P=0.02). There was no difference in predialysis cystatin C concentrations in HDF and high-flux HD groups (P=0.58).

Cystatin C reduction ratio correlated significantly with weekly standard Kt/V (r=0.49, P=0.01) but not single pool Kt/V (r=0.35, P=0.08). Predialysis plasma cystatin C correlated negatively with residual renal urea clearance and GFR (r=-0.51, P=0.01 in both cases).

Predialysis cystatin C concentration correlated significantly with GFR (P=0.01, r=0.51).

Two-Pool Modeling of Cystatin C Clearance

Overall Model Results. Parameter estimates for KNR, Kint, and ECW/Fraction were obtained from iterative modeling for each individual subject. Figure 5 shows combined data from all patients comparing measured and modeled cystatin C levels at each time point throughout the dialysis cycle. Goodness of fit, measured as SD of the difference between the measured and modeled data, had a mean of 0.19 mg/L (SD=0.10), with a range of 0.03–0.42 mg/L. Using the coefficient of determination (r²) to assess goodness of fit, mean r² was 0.89 (SD=0.09), with a range of 0.67–0.99. In the postdialysis period, there was no significant difference between the modeled and observed data using paired t tests (P>0.05). However, in the interdialytic period, the model underestimated cystatin C at 80% and 100% of dialysis duration time points (P<0.001 and P=0.04, respectively), although by only a small magnitude, with mean differences between the data and the model of 0.21 and 0.14 mg/L, respectively.

Iterated Parameter Estimates from Model. Mean iterated KNR was 25.1 ml/min per 1.73 m² body surface area (SD=6.6). The transfer coefficient for the intracellular–extracellular pool barrier (Kint) showed a wide variation between patients, with a median of 63.7 ml/min per 1.73 m² body surface area (interquartile range=29.4–157.1). These clearance parameters are corrected to body surface area to
allow comparison with KR, which is usually body surface area–adjusted. The mean modeled ratio of the extracellular water to total body water (ECWFraction) was 0.255 (SD=0.092).

**Prediction of Postrebound Cystatin C Concentration**

There was no significant correlation between dialysis duration and the magnitude of postdialysis rebound measured by the rise in plasma cystatin C from the end of dialysis to 2 hours postdialysis. The magnitude of the modeled rebound at 2 hours postdialysis was 50.0%±10.9% of the intradialytic fall in cystatin C.

**Discussion**

In the normal population, cystatin C may be superior to creatinine as a single unadjusted marker for predicting GFR (5,32-34), although the finding is not universal (35). Cystatin C–based equations perform as least as well as, if not better than, the Modification of Diet in Renal Disease equation in individuals without substantial renal impairment (3,36-40), although perhaps less well in stage 3 CKD (8). However, in end stage renal failure, its usefulness as a predictor of RRF has not been established. We set out to define the kinetics of cystatin C and study whether it is possible to accurately model cystatin C concentration, which might allow RRF to be estimated from plasma levels.

We have shown that the kinetics of cystatin C are more complex than the kinetics of small molecules, such as urea. Although we were able to accurately model cystatin C concentration using a two-pool iterative model, important model parameters (ECWFraction, $K_{intr}$, and $K_{NR}$) showed large interindividual variance. Mean modeled $K_{NR}$ of 25.1 ml/min per 1.73 m² body surface area was similar to that reported by Sjöström et al. (21), perhaps unsurprisingly, because we used the estimated G from this publication. Levels, however, showed wide variation.

Because predialysis (plateau) concentration of cystatin C is defined by the balance of generation and clearance (renal...
and nonrenal), the wide variation in the ratio of generation rate to nonrenal clearance limits the usefulness of predialysis levels as a marker of RRF in this setting. Assuming that generation rate is relatively stable, then at high levels of GFR, predialysis cystatin C concentration will be largely determined by GFR, whereas at low levels, nonrenal clearance will predominate. In line with this finding, in peritoneal dialysis patients, GFR predicted by a cystatin C–based equation (10) correlated with EDTA clearance but only explained 23% of the variance (41).

Cystatin C clearance during dialysis was of less than that of urea, creatinine, and β2-microglobulin, reflecting its molecular size (13 kD). Cystatin C reduction ratio was greater with high-flow HD than high-flux HD, which has been shown previously (42,43). The rate of cystatin C reduction fell during the HD session, suggesting a lag in transfer to the vascular compartment. Mass removal of cystatin C is greater in the initial phase of dialysis, suggesting that increments in sessional frequency may be a more effective removal strategy than increased duration. We found, as have others, that predialysis cystatin C levels correlate with weekly standard Kt/V but not single pool Kt/V (19,20). This finding probably reflects the contribution of RRF to the former parameter. The relationship has been used by the work by Huang et al. (13), which recently described a method of predicting GFR based on plasma cystatin C levels corrected for standard Kt/V.

Of the solutes studied, the magnitude of the postdialysis rebound was greatest for cystatin C. Unlike the other molecules, cystatin C levels plateaued in the interdialytic period. This finding confirms that predialysis levels are in steady state, in which generation rate is balanced by clearance (renal and nonrenal). The wide interindividual variation in KNR that we have shown implies that predialysis cystatin C concentration will not correlate closely with GFR.

In our two-pool model, the rebound kinetics of cystatin C are predominantly defined by the relative pool sizes of ECW and ICW (determined by ECWFraction) and the mass transfer rate between body compartments (Kint). Both showed wide variance. The apparent relative pool size of the dialyzed extracellular pool in relation to total body water (ECWFraction) was smaller than that predicted from anthropometric equations, which was hypothesized by Tidman et al. (44). Hence, rebound characteristics are not fully explained by re-equilibration of cystatin C from the intracellular to extracellular pool. The reduction in apparent dialyzed pool size implies the presence of one or more pools of sequestered cystatin C. It should be emphasized that ECWFraction represents an apparent pool size derived from modeling rather than a true physiologic parameter.

There is evidence that cystatin C is dimerized and monomerizes as part of the normal cellular processing pathway (45). There are no data on the relative pool size of dimerized cystatin C. It is likely that the assay that we used measured predominantly monomeric cystatin C, although there are no published data. Cystatin C may also be bound to the C4 component of complement (46) and other proteins, which may contribute to the reduction in apparent pool size. In our model, we iterated the relative sizes of the apparent intracellular and extracellular spaces, which will have taken into account the effect of any additional pools. Similar multiple pool effects have been shown for other molecules, such as serum phosphate (25).

Our model assumed also that transfer across the intracellular–extracellular space was a concentration-driven process akin to diffusion, but the process may be more complex and involve active secretion, which is known to occur for cystatin C in the endoplasmic reticulum–lysosome pathway (45,47). Wallin et al. (47) have recently identified a hydrophobic cystatin C region in the cysteine cathepsin binding region of the molecule that seems to be a crucial determinant of cellular uptake. We iterated the mass transfer coefficient (Kint), and, therefore, the parameter is likely to represent the combined effects of diffusion, active transport, and perhaps, other cellular processes that might affect the rebound in cystatin C.

We chose to fix G at 0.124 mg/min per 1.73 m² as previously reported, because it seems to vary little between individuals (11). In the model, it was necessary to constrain either G or KNR to prevent multiple solutions to the model being possible (multiple global minima). By constraining G, we may have overestimated the interindividual variation in nonrenal clearance. The model also assumed that cystatin C generation occurred into the intracellular space and that concentrations were uniform throughout both pools. However, there is evidence that, although cystatin C functions as a housekeeping protein, concentration varies in different body fluid spaces. The dominant type of cysteine protease inhibitors may also vary between tissues (48). Body water was not iterated, because it would have resulted in models with multiple solutions. Similarly, dialyzer clearance was not iterated but included as a constant estimated from in vivo clearance data for β2-microglobulin, a similar-sized middle molecule. Limited published data suggest that cystatin C clearance is approximately 90% of β2-microglobulin clearance for high-flux dialyzers in HD and HDF (26–28). There are extensive in vivo clearance data available for β2-microglobulin (29) that are specific to dialysis modality (high-flux HD or HDF), blood flow, and dialyzer size. However, we acknowledge that more extensive in vivo cystatin C clearance studies are required to confirm this result.

Despite these limitations, this two-pool model is able to accurately model the rebound in cystatin C, with the modeled concentration not differing significantly from the measured concentration at any time point. In the interdialytic period, fit was not quite as good, with modeled levels differing, albeit marginally, from measured levels at the two time points toward the end of the dialysis session. This result may suggest that ECWFraction decreases to the end of dialysis, reflecting release of cystatin C from sequestered pools, perhaps because of monomerization.

In conclusion, plasma cystatin C concentration may be successfully modeled using two-pool kinetics, but it is necessary to iterate the relative sizes of the intracellular and extracellular pools nonphysiologically. This finding implies that the kinetics are substantially more complex than for molecules such as urea and that the incorporation of multiple pools would be required to improve modeling further. Importantly, the rebound in plasma cystatin C is of large magnitude, and the kinetics are difficult to predict. The wide variation in interindividual modeled nonrenal clearance indicates that, at low levels of GFR, single plasma
cystatin C concentrations are unlikely to be useful as an indicator of RRF. Lack of interindividual predictability of modeled parameters indicates that, although cystatin C rebound may be modeled using two pools for one individual, prediction of interdialytic concentration for another individual without formal modeling is not possible, unlike for urea. These findings show the limitations in using single point plasma measures of cystatin C to accurately predict GFR in patients on HD. Cystatin C is cleared by high-flux membranes, and its usefulness as a marker of middle-molecule clearance should be further explored.

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Disclosures
None.

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Supplementary Material

Methods

Hemodialysis and hemodiafiltration procedures

All HD sessions were performed using Fresenius 5008 HD machines and were delivered as either high-flux HD or online HDF. Bicarbonate was used exclusively as a buffer, and ultrapure water used for dialysis and for replacement fluid in the case of online HDF. In 22 out of 24 patients Fresenius FX class high-flux dialysers were used (FX60, FX80, FX100). In the remaining two cases, dialysers used were Arylane H9 and Gambro Nephral 500ST. All patients were dialysed using an incremental HD algorithm whereby the target Kt/V was 1.2 and was considered the sum of dialysis two-pool eKt/V and the equivalent Kt/V provided by RRF, as described previously¹. For online HDF, the HDF fraction was 0.35 of the plasma flow rate and target HDF volume was 40% of the Watson volume. Vascular access was via tunnelled vascular catheters or arteriovenous fistulas. Ultrafiltration was performed to achieve a clinically assessed dry weight.

Plasma sample analysis

Blood samples were collected in lithium heparin bottles (Vacuette®, Greiner-Bio, Frickenhausen, Germany), centrifuged, and stored at -80° C until the time of analysis. All samples were analysed for urea, creatinine, β2-microglobulin and cystatin C according to the manufacturer specifications in a Clinical Pathology Accreditation approved laboratory. Urea, creatinine and β2-microglobulin were measured using an Olympus AU640 (Beckman-Coulter, High Wycombe, UK). The specific methodology for these include colorimetric analysis (modified Jaffé method)
for creatinine and immune-turbidimetric for β2-microglobulin. The between-day assay coefficient of variation for β2-microglobulin was 4.8% at a mean concentration of 6.22 mg/l and assay calibration was performed before measuring all samples. External quality assurance was also performed routinely for this assay throughout the period of analysis and showed acceptable performance. Cystatin C was measured on the IMMAGE®800 (Beckman-Coulter, High-Wycombe, UK) immunoassay analyser using DakoCytomation cystatin C reagent (Dako Ltd, Cambridgeshire, UK) based on the non-competitive rate particle-enhanced nephelometric technology. Repeated analysis of patient samples showed <3 % inter-assay variation.

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