Fibroblast Growth Factor 23 and Cardiovascular Mortality after Kidney Transplantation

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Abstract

Background and objectives Circulating fibroblast growth factor 23 (FGF23) is associated with adverse cardiovascular outcomes in CKD. Whether FGF23 predicts cardiovascular mortality after kidney transplantation, independent of measures of mineral metabolism and cardiovascular risk factors, is unknown.

Design, setting, participants, & measurements The association between plasma C-terminal FGF23 and cardiovascular mortality was analyzed in a single-center prospective cohort of 593 stable kidney transplant recipients (mean age ± SD, 52±12 years; 54% male; estimated GFR, 47±16 ml/min per 1.73 m²), at a median of 6.1 (interquartile range, 2.7–11.7) years after transplantation. Multivariate Cox regression models were built, adjusting for measures of renal function and mineral metabolism; Framingham risk factors; the left ventricular wall strain markers midregional fragment of pro–A-type natriuretic peptide (MR-proANP) and N-terminal-pro brain natriuretic peptide (NT-proBNP); and copeptin, the stable C-terminal portion of the precursor of vasopressin.

Results In multivariate linear regression analysis, MR-proANP (β =0.20, P<0.001), NT-proBNP (β =0.18, P<0.001), and copeptin (β =0.26, P<0.001) were independently associated with FGF23. During follow-up for 7.0 (interquartile range, 6.2–7.5) years, 128 patients (22%) died, of whom 66 (11%) died due to cardiovascular disease; 54 (9%) had graft failure. FGF23 was associated with an higher risk of cardiovascular mortality in a fully adjusted multivariate Cox regression model (hazard ratio [HR], 1.88 [95% confidence interval (CI), 1.11 to 3.19]; P=0.02). FGF23 was also independently associated with all-cause mortality (full model HR, 1.86 [95% CI, 1.27 to 2.73]; P=0.001). Net reclassification improved for both cardiovascular mortality (HR, 0.07 [95% CI, 0.01 to 0.14]; P<0.05) and all-cause mortality (HR, 0.11 [95% CI, 0.05 to 0.18]; P<0.001).

Conclusions Plasma FGF23 is independently associated with cardiovascular and all-cause mortality after kidney transplantation. The association remained significant after adjustment for measures of mineral metabolism and cardiovascular risk factors.

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Introduction

Kidney transplantation is the treatment of choice for patients with ESRD. As a result of improved surgical techniques, immunosuppressive therapies, and prevention of opportunistic infections, the short-term prognosis of kidney transplant recipients in particular has improved dramatically over the past decades. However, the risk of premature death due to cardiovascular disease remains greatly increased in kidney transplant recipients compared with the general population (1).

Deregulation of calcium/phosphate metabolism is common in CKD and in kidney transplant recipients with impaired renal function (2). In patients with CKD, circulating levels of fibroblast growth factor 23 (FGF23), parathyroid hormone (PTH), and phosphate have been identified as independent risk factors for cardiovascular disease and all-cause mortality (3–9). Experimental studies demonstrated that FGF23 may be directly involved in the development of left ventricular hypertrophy (LVH) (10). Whether FGF23 predicts cardiovascular mortality in renal transplant recipients is unknown.

Our primary hypothesis was that plasma FGF23 is a risk factor for cardiovascular mortality in renal transplant recipients, independent of Framingham risk factors (recipient age and sex, systolic BP, antihypertensive treatment use, smoking status, diabetes mellitus, plasma total cholesterol, and HDL cholesterol), known correlates of FGF23 (including estimated GFR [eGFR] and proteinuria [11]), and factors related to phosphate metabolism. We also addressed whether FGF23 is associated with the left ventricular wall strain markers midregional fragment of pro–A-type natriuretic peptide (ANP) and N-terminal-pro brain natriuretic peptide (NT-proBNP) and with copeptin, the stable C-terminal portion of the precursor of vasopressin (12), and whether these cardiac markers

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Materials and Methods

Study Population

In this prospective cohort study, all renal transplant recipients with a graft functioning for at least 1 year who visited our outpatient clinic (University Medical Center Groningen, The Netherlands) between August 2001 and July 2003 were eligible to participate. This study was a post hoc analysis of a previously published study (13); plasma samples from 593 patients were available. All participants provided written informed consent. Patients were enrolled at a median of 6.1 (interguartile range [IQR], 2.7-11.7) years after transplantation. Baseline visits were postponed if signs of infection were present. Patients diagnosed with cancer other than cured skin cancer were not eligible. The institutional review board approved the study protocol (METc 01/039), which was in adherence with the Declaration of Helsinki. The clinical and research activities being reported are consistent with the Principles of the Declaration of Istanbul as outlined in the Declaration of Istanbul on Organ Trafficking and Transplant Tourism.

Endpoints

The major outcome of the study was cardiovascular mortality. All-cause mortality and death-censored graft failure were secondary outcomes. Adequate collection of up-to-date data on events and mortality was ensured by our continuous surveillance system; general practitioners or referring nephrologists were contacted in case the current status of a patient was unknown. Cause of death was obtained by linking the number of the death certificate to the primary cause of death as coded by a physician, according to the International Classification of Diseases, Ninth Revision (ICD-9). Cardiovascular death was defined as ICD-9 codes 410–447 (14). Follow-up was completed until May 19, 2009. There was no loss to follow-up.

Patient Characteristics and Laboratory Measurements

Relevant transplant characteristics were taken from a database containing information on all renal transplantations performed at our center since 1968. Standard immunosuppressive treatment regimens are published previously (13). Current medication, including active vitamin D use (alfacalcidol or calcitriol); presence of diabetes mellitus; and cardiovascular history were extracted from the medical record. Diabetes mellitus was defined according to the American Diabetes Association guidelines: fasting plasma glucose $\geq 126 \text{ mg/dl}$ or use of antidiabetic medication (15). Cardiovascular history was defined as a history of myocardial infarction, percutaneous transluminal angioplasty or stenting of coronary or peripheral arteries, bypass operation of coronary or peripheral arteries, intermittent claudication, amputation for vascular reasons, transient ischemic attack, or ischemic cerebrovascular accident. BP was measured as described previously (13). Smoking status was recorded with a self-report questionnaire.

Upon entry in the cohort (baseline), blood was drawn after an 8- to 12-hour overnight fasting period. EDTA

plasma samples were stored at -80°C until assessment of biochemical measures for this study. Plasma C-terminal FGF23 levels were determined by sandwich ELISA (Immutopics, San Clemente, CA), with intra-assay and interassay coefficients of variation of <5% and <16%, respectively (16). Plasma creatinine concentrations were determined using a modified version of the Jaffe method (MEGA AU 510, Merck Diagnostic, Darmstadt, Germany). PTH and 1,25(OH)₂ vitamin D were measured in EDTA plasma using radioimmunoassay, and 25(OH)-vitamin D levels were determined by isotope dilution-online solid phase extraction liquid chromatography-tandem mass spectrometry (17). NT-proBNP levels were measured by immunoassay on an ELECSYS2010 instrument (ELECSYS proBNP, Roche Diagnostics, Germany). MR-proANP was measured with a sandwich immunoassay (MR-proANP LIA; B.R.A.H.M.S) (18). Plasma copeptin was measured using a sandwich immunoassay (B.R.A.H.M.S GmbH/Thermo Fisher Scientific, Hennigsdorf/Berlin, Germany) (19). Serum albumin, calcium, cholesterol, C-reactive protein, glucose, hemoglobin, and phosphate and urinary phosphate, sodium, total protein, and urea were determined by routine laboratory measurements. We calculated eGFR using the CKD-Epidemiology Collaboration equation (20).

Statistical Analyses

Variable distribution was tested with histograms and probability plots. Normally distributed variables are presented as mean \pm SD, and skewed variables as median (interquartile range). Baseline characteristics upon entry into the cohort were compared between tertiles of FGF23 using one-way ANOVA, Kruskal-Wallis test, and chi-squared tests as appropriate. Skewed data were natural log-transformed for correlation analysis, Cox regression, and backward and forward linear analysis.

The associations between (log-transformed) FGF23 levels and MR-proANP, NT-proBNP, and copeptin were assessed by Pearson correlation analysis. Backward linear regression was used to identify correlates of plasma FGF23 levels. The following covariates were tested: age, sex, donor type (deceased or living), history of one or more acute rejection episodes, cold ischemia time, waist circumference, cardiovascular history, Framingham risk score factors, eGFR, proteinuria, serum phosphate, 24-hour urinary phosphate excretion (representing phosphate intake), 24-hour urinary urea excretion (representing protein intake [21]), plasma 25 (OH)-vitamin D, 1,25(OH)₂ vitamin D, PTH, C-reactive protein, albumin, hemoglobin, MR-proANP, NT-proBNP, copeptin levels, use of angiotensin-converting enzyme inhibitor or angiotensin receptor blocker (given potential interactions between the renin-angiotensin-aldosterone system and FGF23 [22]), use of active vitamin D, and number of antihypertensive drugs. Variables significantly associated with FGF23 levels using backward linear regression analysis were subsequently tested in a forward linear regression model. Variables that were significant in this model were considered independent correlates of FGF23 levels.

Risk ratios and conditional maximum likelihood estimates of the rate ratio for cardiovascular and all-cause mortality and graft failure were calculated per tertile of

Table 1. Clinical characteristics of the co	ohort				
			Plasma FGF2	3 Tertiles	
Variable	All Patients (<i>n</i> =593)	Tertile 1 (<i>n</i> =197)	Tertile 2 (<i>n</i> =198)	Tertile 3 (<i>n</i> =198)	P Value
Plasma C-terminal FGF23 (RU/ml) Age (yr) Men (%) Whites (%)	$\begin{array}{c} 140 \ (95-219) \\ 52 \pm 12 \\ 54 \\ 95 \end{array}$	$\begin{array}{c} 83 \ (68-96) \\ 50 \pm 13 \\ 56 \\ 94 \end{array}$	141 (124–162) 52±12 51 94	311 (220–567) 53±11 55 95	$< 0.001 \\ 0.04 \\ 0.62 \\ 0.41$
Cause of kidney disease (%) Primary GN GN due to vasculitis/	28 6	26 9	30 5	29 6	0.75
Tubulointerstitial nephritis/	16	20	15	14	
Polycystic kidney disease Hypertension Diabetes	18 6 3	16 7 4	18 5 3	20 5 3	
Other/unknown Dialysis duration (mo) Deceased donor transplant (%)	23 27 (13–48) 86	18 24 (13–43) 78	24 31 (14–48) 92	23 26 (13–52) 91	0.40 < 0.001
Cold ischemia time (h) Total warm ischemia time (min) Total HLA mismatches (<i>n</i>) History of acute rejection (%)	22 (15–27) 35 (30–45) 1 (0–2) 45	20 (7–27) 35 (30–45) 1 (0–2) 39	22 (16–26) 35 (30–45) 1 (0–2) 44	22 (16–29) 36 (30–45) 1 (0–2) 52	0.04 0.92 0.41 0.04
Transplant vintage (yr) Waist circumference (cm) Systolic BP (mmHg) Diastolic BP (mmHg)	$\begin{array}{c} 6.1 \ (2.7 - 11.7) \\ 97 \pm 14 \\ 153 \pm 23 \\ 90 \pm 10 \end{array}$	$5.6 (2.4-11.1) 94\pm13 148\pm21 89\pm10$	$5.8 (3.0-11.5) 97 \pm 14 153 \pm 22 89 \pm 9$	$7.2 (3.5-12.2) 100\pm13 157\pm25 91\pm10$	0.06 < 0.001 < 0.001 < 0.001 < 0.001
Current smoker (%) Current diabetes (%) Laboratory results	22 18	18 19	18 17	30 18	<0.001 0.88
Creatinine (mg/dl) Albumin (g/dl) Calcium (mg/dl)	$\begin{array}{c} 1.5 \ (1.3 - 1.9) \\ 4.1 \pm 0.3 \\ 9.6 \pm 0.6 \end{array}$	$\begin{array}{c} 1.4 \ (1.1 - 1.6) \\ 4.1 \pm 0.3 \\ 9.5 \pm 0.6 \end{array}$	$\begin{array}{c} 1.5 \ (1.3 - 1.7) \\ 4.1 \pm 0.4 \\ 9.6 \pm 0.7 \end{array}$	$\begin{array}{c} 1.9 \; (1.4 - 2.5) \\ 4.0 \pm 0.3 \\ 9.6 \pm 0.7 \end{array}$	<0.001 <0.001 0.20
Total cholesterol (mg/dl) HDL cholesterol (mg/dl) LDL cholesterol (mg/dl)	218±42 42±12 137±39	214 ± 35 45 ± 13 137 ± 31 180(02,410)	219±38 42±12 136±39	$221 \pm 51 \\ 40 \pm 12 \\ 138 \pm 46 \\ 457 (240, 1517)$	0.13 <0.001 0.84
MI-probINP (pg/ml) MR-proANP (pmol/L) Copeptin (pmol/L) C-reactive protein (mg/L)	$\begin{array}{c} 299 (131-672) \\ 164 (103-275) \\ 9.3 (5.0-19.5) \\ 2.1 (0.8-4.9) \end{array}$	180(92-419) 126(87-188) 6.5(3.8-10.4) 1.7(0.6-3.7)	285 (130–367) 153 (100–231) 8.6 (4.7–17.4) 2.1 (0.8–4.5)	457 (240–1517) 257 (158–390) 16.1 (7.7–30.0) 3.0 (1.2–7.8)	<0.001 <0.001 <0.001 <0.001
Glucose (mg/dl) Phosphate (mg/dl) Parathyroid hormone (pg/ml)	87±23 3.3±0.7 86 (57–131)	86±22 3.1±0.6 72 (51–112) 22+10	87±26 3.2±0.6 88 (61–121)	88±20 3.6±0.7 101 (68–193) 22+10	0.59 < 0.001 < 0.001 < 0.001
1,25-OH $_2$ vitamin D (ng/ml) Hemoglobin (g/dl) Estimated GFR (CKD-EPI)	21 ± 9 42 ± 18 13.7 ± 1.5 47 ± 16	22 ± 10 47 ± 17 14.2 ± 1.3 55 ± 13	21 ± 0 42 ± 19 13.8 ± 1.3 48 ± 14	22 ± 10 35 ± 15 13.1 ± 1.7 37 ± 15	<0.49 <0.001 <0.001 <0.001
(ml/min per 1.73 m ²) Measurements in 24-hour urine Sodium excretion (mEq/24 hr) Phosphate excretion (g/24 hr)	140 ± 68 2 3+0 8	139 ± 68 2 3+0 9	138 ± 61 2 4 ± 0 8	143 ± 75 2 2+0 8	0.66
Proteinuria (g/24 hr) Urea excretion (g/24 hr) Medication	0.2 (0.0–0.5) 10.5±3.2	0.2 (0.0–0.3) 10.6±3.1	0.2 (0.0–0.4) 10.9±3.2	0.3 (0.2–0.8) 10.0±3.2	<0.001 0.03
Calcium supplements (%) Active vitamin D (alfacalcidol or calcitriol) (%)	7 9	5 8	6 5	10 14	0.07 0.02
ACE inhibitor or ARB (%) Prednisone dose (mg/d)	34 10 (7.5–10)	30 10 (7.5–10)	34 10 (7.5–10)	39 10 (7.5–10)	0.17 0.58

Table 1. (Continued)							
		Plasma FGF23 Tertiles					
Variable	(<i>n</i> =593)	Tertile 1 (<i>n</i> =197)	Tertile 2 (<i>n</i> =198)	Tertile 3 (<i>n</i> =198)	P Value		
Calcineurin inhibitor (%)	78	77	83	75	0.12		
Azathioprine (%)	34	29	34	38	0.17		
Mycophenolate mofetil (%)	40	49	41	31	< 0.001		
Sirolimus (%)	2	2	2	2	0.88		

Values expressed with a plus/minus sign are the mean ± SD; values with parentheses are median and interquartile range. All variables were determined at enrollment except sex, race, cause of the original kidney disease, cold ischemia time, total warm ischemia time, and total number of HLA mismatches, which were assessed at time of transplantation. FGF23, fibroblast growth factor 23; NT-proBNP, N-terminal-pro brain natriuretic peptide; MR-proANP, pro–A-type natriuretic peptide; CKD-EPI, CKD-Epidemiology Collaboration; ACE, angiotensin-converting enzyme; ARB, angiotensin receptor blocker.

FGF23. Confidence intervals were calculated using the Fisher exact test. To further analyze the association between FGF23 and outcomes, tertiles of FGF23 were plotted in Kaplan-Meier curves with a log-rank test. Subsequently, multivariate Cox regression models were built to assess the association between FGF23 and outcomes, adjusted for potential confounders. We tested for effect modifications by invoking multiplicative interaction terms. Sensitivity analyses were performed in a predefined subgroup of patients with an eGFR between 30 and 90 ml/min per 1.73 m² (n=493).

To determine the goodness of fit of the predictive models, the Akaike information criterion (AIC) was used. AIC is a statistical estimate of the trade-off between the likelihood of a model against its complexity. A lower AIC value indicates a better model fit. Furthermore, discrimination, or the ability of the risk prediction models to distinguish those with an event (*i.e.*, cardiovascular or allcause mortality or graft failure) from those who do not, was evaluated with the Harrell C-index. The C-index is analogous to the area under the receiver-operating characteristic curve, for which larger values indicate better discrimination.

We assessed net reclassification improvement (NRI) for predefined risk categories of cardiovascular and all-cause mortality (<5%, 5%–10%, and >10%) and calculated integrated discrimination improvement (IDI) (23). The NRI provides reclassification tables constructed separately for study participants with and without events and quantifies the correct movement between categories: upwards for events and downward for nonevents.

Proportionality assumptions were tested using Schoenfeld tests and log-minus-log survival plots. All statistical analyses were performed using SPSS software, version 16.0 for Windows (IBM, Armonk, NY); Stata/SE software, version 11.0 for Windows (College Station, TX); and OpenEpi (openepi.com). *P* values <0.05 were considered to represent statistically significant differences.

Results

Patient Characteristics

Patient characteristics are summarized in Table 1. Overall, FGF23 levels were 140 (IQR, 95–219) RU/ml, and eGFR was 47 ± 16 ml/min per 1.73 m². None of the patients used dietary phosphate binders. Patients in the highest FGF23 tertile were older; were more likely to have received a graft from a deceased donor, to have had an acute rejection episode, and to be a smoker; had a longer cold ischemia time; had a higher waist circumference and systolic and diastolic BP; had worse renal function; had lower serum albumin, HDL cholesterol, $1,25(OH)_2$ vitamin D, and hemoglobin; had higher serum phosphate, NT-proBNP, MR-proANP, copeptin, PTH, and CRP levels; and had more proteinuria.

FGF23 Is Independently Associated with Cardiac Markers

Plasma FGF23 levels were associated with NT-proBNP (r^2 =0.12; P<0.001), MR-proANP (r^2 =0.18; P<0.001), and copeptin (r^2 =0.18; P<0.001) levels in univariate analysis (Figure 1). Using backward and forward multivariate linear regression models, we found that copeptin was strongly and independently associated with plasma FGF23 levels (Table 2). Other independent correlates of FGF23 levels were eGFR, serum phosphate, hemoglobin, and donor type (living or deceased). Results were highly similar when, instead of copeptin, MR-proANP (coefficient, 0.20 [95% CI, 0.12 to 0.28]; P<0.001) or NT-proBNP (coefficient, 0.18 [95% CI, 0.11 to 0.25]; P<0.001) were forced into the final forward regression model.

Baseline Plasma FGF23 and Outcomes

During a median follow-up of 7.0 (IQR, 6.2–7.5) years, 128 patients (22%) died, 66 (11%) of cardiovascular disease; 54 (9%) had graft failure. Kaplan-Meier survival plots demonstrated a higher risk of cardiovascular mortality, all-cause mortality, and graft failure per increasing tertile of plasma FGF23 (Figure 2). Risk ratios for cardiovascular mortality, all-cause mortality, and graft failure were significantly increased in the highest FGF23 tertile (Table 3).

In Cox regression models, the highest tertile of FGF23 was consistently associated with a higher risk of cardiovascular mortality after adjustment for Framingham risk factors, history of cardiovascular disease, measures of renal function, phosphate metabolism, and cardiac wall stress (Table 4). To assess whether the association between FGF23 and cardiovascular mortality was influenced by



Figure 1. | Scatter diagrams illustrating the associations between fibroblast growth factor 23 (FGF23) and (A) N-terminal-pro brain natriuretic peptide (NT-proBNP), (B) pro–A-type natriuretic peptide (MR-proANP), and (C) copeptin.

other factors, interaction analyses were performed. We found no evidence for interaction in the association between FGF23 and cardiovascular mortality by eGFR (P for interaction=0.51), proteinuria (P=0.62), 24-hour urinary phosphate excretion (P=0.50), fractional phosphate

Table 2.Multivariate linear regression analysis for correlatesof plasma fibroblast growth factor 23 levels

Variable	Coefficient for FGF23 (95% CI)	P Value
eGFR (CKD-EPI) Ln copeptin Serum phosphate Hemoglobin Donor type (0=living; 1=deceased)	-0.32 (-0.41 to -0.23) 0.26 (0.17 to 0.35) 0.15 (0.07 to 0.23) -0.14 (-0.22 to -0.06) -0.10 (-0.18 to -0.03)	<0.001 <0.001 <0.001 <0.001 0.004

Excluded from the model: age, sex, history of one or more acute rejection episodes, cold ischemia time, waist circumference, cardiovascular history, systolic and diastolic BP, smoking status, diabetes mellitus, total cholesterol, proteinuria, 24-urinary phosphate and urea excretion, plasma 25(OH)-vitamin D, 1,25 (OH)₂ vitamin D, C-reactive protein, albumin, parathyroid hormone, use of angiotensin-converting enzyme inhibitor or angiotensin receptor blocker, use of active vitamin D, and total number of antihypertensive drugs. Coefficients are standardized β values. Model fit: R^2 =0.41. FGF23, fibroblast growth factor 23; CI, confidence interval; eGFR, estimated GFR; CKD-EPI, CKD-Epidemiology Collaboration; Ln; natural log.

excretion (P=0.87), or any Framingham risk factor. In addition, no significant interaction by NT-proBNP (P for interaction=0.26), MR-proANP (P=0.32), or copeptin (P=0.19) was observed. NT-proBNP and MR-ANP but not copeptin were independently associated with cardiovascular mortality in the full model (data not shown). The final models provided the best fit (lowest AICs); all models displayed good discrimination as suggested by relatively high Harrell C-indices (Table 4). As a sensitivity analysis, full multivariate Cox regression models were repeated in a subgroup of patients with an eGFR between 30 and 90 ml/min per 1.73 m² (n=493). In this subgroup, plasma FGF23 remained an independent risk factor for cardiovascular mortality (full model hazard ratio [HR], 2.43 [95% CI, 1.19 to 4.93] per 1 SD of natural log [ln]FGF23; P=0.01) and all-cause mortality (HR, 2.12 [95% CI, 1.29 to 3.49] per 1 SD of lnFGF23; P=0.003).

Higher FGF23 levels were also associated with a higher risk of all-cause mortality, both when analyzed per tertile and as continuous variable (Table 4). Higher tertiles of FGF23 were also associated with a higher risk of deathcensored graft failure in crude analyses, but significance was lost after adjustment for eGFR and proteinuria (Table 4). There was no significant interaction between the risk of graft failure and cardiovascular mortality (P=0.38).

Comparative Analyses

To compare the performance of FGF23 with other measures of mineral metabolism (fractional phosphate excretion, PTH, phosphate, 25-OH vitamin D or 1,25[OH]₂ vitamin D) as individual predictors of cardiovascular mortality, separate Cox regression analyses were performed for each of the six exposures, adjusted for age, sex, cardiovascular history, eGFR, proteinuria, and Framingham risk factors. As indicated in Figure 3, FGF23 (highest tertile HR, 3.45 [95% CI,



Figure 2. | Kaplan-Meier survival plots of fibroblast growth factor 23 (FGF23) tertiles and cardiovascular mortality. Higher FGF23 levels are associated with an increased incidence of cardiovascular mortality.

1.57 to 7.58]; *P*=0.001) but not the other exposures were strongly associated with cardiovascular mortality.

NRI and IDI

The NRI was calculated to assess the putative clinical utility of FGF23 as a marker of (cardiovascular) mortality in addition to Framingham risk factors. Reclassification improved mainly in the patients who survived, where adding FGF23 to the model resulted in reclassification into a lowerrisk category (Table 5). Overall, net reclassification improved for both cardiovascular mortality (0.07 [95% CI, 0.01 to 0.14]; P<0.05]) and all-cause mortality (0.11 [95% CI, 0.05 to 0.18]; P<0.001). Accordingly, the IDI, which does not rely on prespecified risk categories but represents a continuous measure, was 0.03 (95% CI, 0.01 to 0.05; P=0.004) for cardiovascular mortality and 0.05 (95% CI, 0.03 to 0.07; P<0.001) for all-cause mortality.

Discussion

This study identified high plasma FGF23 levels as an independent risk factor for cardiovascular mortality in

kidney transplant recipients. The association was consistent in regression models adjusted for renal function, measures of mineral metabolism, and cardiovascular risk factors. The significantly improved NRI suggests that FGF23 levels may have an additional value to Framingham risk factors to predict the risk of cardiovascular mortality in renal transplant recipients.

Our findings are in line with previous data linking high FGF23 levels with incident cardiovascular disease and mortality in the CKD population (3,24,25), and with a higher risk of all-cause mortality after kidney transplantation (26). Furthermore, FGF23 levels have been associated with established cardiovascular risk factors and with a higher risk of cardiovascular events in the general population (27,28). Although patients in the highest FGF23 tertile were characterized by an overall high cardiovascular risk profile, the associations between FGF23 and (cardiovascular and all-cause) mortality remained significant after adjustment for several major cardiovascular risk markers, including Framingham risk factors, suggesting that FGF23 is a strong and independent risk marker. The potential clinical relevance of FGF23 in addition to established risk

X7 1.1.			Plasma FGF23 Tertiles			
Variable	All Patients	Tertile 1	Tertile 2	Tertile 3	P value	
Cardiovascular mortality						
Events/patient-years	66/3722	11/1343	17/1260	38/1119		
Incidence rate/1000 person-years	17.7	8.2	13.5	34.0		
Risk ratio (95% CI)		Reference	1.65 (0.73 to 3.89)	4.15 (2.08 to 8.99)	< 0.001	
All-cause mortality			· · · · ·			
Events/patient-years	128/3722	24/1343	33/1260	71/1119		
Incidence rate/1000	34.4	17.9	26.2	63.4		
person-years Risk ratio (95% CI)		Reference	1 47 (0 84 to 2 59)	3 55 (2 21 to 5 90)	< 0.001	
Graft failure		Reference	1.17 (0.01 to 2.07)	0.00 (2.21 to 0.90)	<0.001	
Events/patient-years	54/3722	8/1343	8/1260	38/1119		
Incidence rate/1000	14.5	6.0	6.3	34.0		
person-years						
Risk ratio (95% CI)		Reference	1.07 (0.35 to 3.26)	5.70 (2.62 to 14.15)	< 0.001	

^aFor trend.

markers is further supported by improved reclassification (NRI and IDI). Furthermore, FGF23, but not serum phosphate, PTH, or vitamin D, was independently associated with cardiovascular mortality (Figure 3), suggesting a specific role for FGF23. The observed association between elevated FGF23 levels and a higher cardiovascular risk initially positioned FGF23 as a biomarker of phosphate toxicity. More recently, animal studies demonstrated that FGF23 has "off-target" effects, directly contributing to the development of LVH (10). We found that FGF23 was independently associated with MR-proANP and NTproBNP, both markers of left ventricular wall strain, and with copeptin, a stable peptide derived from the vasopressin precursor (29). All three cardiac markers are used as markers of heart failure (30). Although these associations are in line with a potential role for FGF23 in LVH, we could not demonstrate modulation of the association between FGF23 and cardiovascular mortality by these cardiac markers. Furthermore, only a small amount of FGF23 was explained by copeptin, MR-proANP, and NT-proBNP (Figure 1), suggesting that FGF23 is independently related to cardiovascular and all-cause mortality.

In a previous study, the association between FGF23 and cardiovascular events was modified by sex (31). We could not confirm this effect modulation, which could be explained by differences in renal function between the Heart and Soul study (eGFR, 71±23 ml/min per 1.73 m²) and our cohort (eGFR, 47±16 ml/min per 1.73 m²). Data from 24-hour urine collections allowed us to include phosphate intake (assessed by 24-hour urinary phosphate excretion) (32) and excretion (fractional phosphate excretion) in our analyses. Neither measure interacted with the association between FGF23 levels and cardiovascular mortality.

FGF23 was also associated with a higher risk of graft failure in univariate analysis, in line with a previous study (26). However, the association was no longer significant after adjustment for eGFR and proteinuria in addition to known risk factors for graft failure (Table 3). We hypothesize that renal function after transplantation has a stronger effect on the risk of graft failure than on the risk of mortality (33,34).

Possible limitations of our study include its post hoc nature and the use of ICD-9 codes rather than an independent adjudication committee to define the endpoint cardiovascular mortality. Although we adjusted for several confounders, including measures of renal function, which appeared to be the most important determinant of FGF23 levels, the possibility of residual confounding cannot be fully excluded because not all determinants of FGF23 are currently known. The sensitivity analyses in patients with an eGFR of 30-90 ml/min per 1.73 m² reduces the risk of confounding by renal function. The fact that diabetic and hypertensive nephropathy were underrepresented in our cohort, and that our cohort consisted almost entirely of white patients, limits the generalizability of our study. Because FGF23 was measured only at a single time point in this cohort, we could not perform a repeatedmeasures analysis. Prevalent cohorts such as this may suffer from survivorship biases; these generally lead to an underestimation of the hazard ratio (35). Using a relatively large number of covariates in the final Cox regression model might result in overfitting the model; however, the stepwise Cox models and the low AIC in the final model suggest that overfitting was no issue in our analyses. Finally, we did not have extensive structural or functional cardiac data (e.g., from echocardiography) available to further explore the mechanisms behind the observed association between FGF23 and cardiovascular mortality. Strong points of this study, on the other hand, are the use of 24-hour urine collections, which enabled us to adjust for proteinuria and phosphate excretion, the availability of multiple markers of cardiovascular risk, and the complete follow-up.

In conclusion, to our knowledge this is the first study to identify FGF23 as an independent risk factor for

Table 4. Cox regression analyse	es for cardiovascu	ılar and all-cause mortalit	y and graft failure per fibr	oblast growth fi	actor 23 tertile or per 1 S	D of natural lo	g fibroblast	growth factor 23
¥7		Plasma FGl	F23 Tertiles			Natural Log	FGF23	
v агтарте	Tertile 1	Tertile 2	Tertile 3	P Value ^a	Per 1 SD	P Value	AIC	Harrell C-Index
Cardiovascular mortality, HR (95% CI)								
Model 1 Model 2	Reference Reference	1.66 (0.78 to 3.54)	4.24 (2.17 to 8.29) 4.02 (1.90 to 8.50)	<0.001	1.97 (1.52 to 2.55) 1 98 (1 44 to 2 74)	<0.001	645 648	0.706
Model 3	Reference	1.39 (0.55 to 3.50)	4.20 (1.90 to 9.81)	<0.001	1.93 (1.33 to 2.80)	0.001	800	0.801
Model 4	Reference	1.59 (0.58 to 4.14)	4.06(1.55 to 9.98)	0.003	2.27 (1.37 to 3.75)	0.001	479	0.810
Model 5	Reference	1.86 (0.67 to 5.18)	2.96 (1.06 to 8.26)	0.04	1.88 (1.11 to 3.19)	0.02	467	0.843
All-cause mortality, HR (95% CI)								
Model 1	Reference	1.47 (0.87 to 2.48)	3.64 (2.29 to 5.78)	< 0.001	1.99 (1.65 to 2.39)	< 0.001	1245	0.694
Model 2	Reference	1.36 (0.80 to 2.32)	2.94 (1.73 to 4.98)	< 0.001	1.86(1.46 to 2.36)	< 0.001	1237	0.693
Model 3	Reference	1.03 (0.55 to 1.93)	3.15(1.69 to 5.85)	< 0.001	1.93 (1.45 to 2.56)	< 0.001	1542	0.768
Model 4	Reference	1.06 (0.53 to 2.11)	2.63 (1.32 to 5.24)	0.004	2.18 (1.50 to 3.18)	< 0.001	869	0.778
Model 5	Reference	1.17 (0.58 to 2.36)	2.23 (1.11 to 4.49)	0.02	1.86(1.27 to 2.73)	0.001	858	0.799
Graft failure, HR (95% CI)								
Model 1	Reference	1.06 (0.40 to 2.83)	6.29 (2.92 to 13.44)	< 0.001	3.37 (2.56 to 4.42)	< 0.001	515	0.762
Model 2	Reference	0.54 (0.20 to 1.46)	1.15 (0.47 to 2.84)	0.41	1.49 (0.99 to 2.24)	0.06	463	0.861
Model 3	Reference	0.53 (0.18 to 1.57)	0.80 (0.29 to 2.24)	0.99	1.15 (0.73 to 1.82)	0.55	540	0.885
Model 4	Reference	0.35 (0.10 to 1.22)	0.39 (0.12 to 1.30)	0.25	$1.00\ (0.59\ to\ 1.71)$	0.99	327	0.907
Model 5	Reference	0.32 (0.09 to 1.21)	0.37 (0.11 to 1.32)	0.25	1.03 (0.59 to 1.78)	0.93	331	0.909
Model 1: crude model; model 2: Framinoham rick factore (recinie	adjusted for estin	nated GFR (CKD-Epidem scholic RP antibunertensi	iology Collaboration form	ula) and proteir status diabetes	uria; model 3: model 2 + mellitus, masma total cho	adjusted for car lecterol and E	trdiovascula TDI choleste	r disease history,
vintage; model 4: model 3 + adju	sted for serum ph	tosphate, 1,25(OH) ₂ -vitan	nin D, parathyroid hormon	e levels, and ac	ive vitamin D use; model	5: model 4 + ac	djusted for N	L-terminal-pro brain
natriuretic peptide, pro-A-type 1 ^a P for linear trend.	aatriuretic peptid	e, and copeptin. HR, haza	ard ratio; CI, confidence int	terval; AIC, Ak	aike information criterion			



Figure 3. | Comparative analysis of fibroblast growth factor 23 (FGF23), fractional phosphate excretion (FE PO4), parathyroid hormone, phosphate (P), 25(OH)-vitamin D, or 1,25(OH)₂ vitamin D as independent risk factors for cardiovascular (CV) mortality. The model was adjusted for age, sex, cardiovascular history, estimated GFR, proteinuria, and Framingham risk factors. For each exposure, the lowest tertile served as the reference group (R).

cardiovascular mortality after kidney transplantation. Although it may be relevant to consider therapies reducing FGF23 levels (36) to improve cardiovascular outcomes, it should be kept in mind that high FGF23 levels may serve an important physiologic goal, namely to keep phosphate balance. A recent study demonstrated that specific FGF23 blockade with a neutralizing antibody did reduce secondary hyperparathyroidism but increased serum phosphate, aortic calcification, and mortality (37). Whether specific reduction of FGF23 or rather reducing phosphate load may improve cardiovascular prognosis after kidney transplantation should be addressed in future prospective studies.

Table 5. Reclassification based on fibroblast growth factor	23 levels			
With and ECEO2		$T_{abal}(u)$		
without FGF25	<5%	5%-10%	>10%	10tal (<i>n</i>)
Estimated risk of cardiovascular mortality ^a				
Patients with cardiovascular mortality				
<5%	2	1 ^b		3
5%-10%		6		6
>10%		2^{c}	55	57
Total	2	9	55	66
Patients without cardiovascular mortality				
<5%	186	12 ^c	1 ^c	199
5%-10%	46 ^b	75	15 ^c	136
>10%		27 ^b	15	178
Total	232	114	167	513
Estimated risk of all-cause mortality ^a				
Patients with all-cause mortality			,	
<5%	1	1 ^b	1 ^b	3
5%-10%	3°	3	2 ^b	8
>10%		3°	114	117
Total	4	7	117	128
Patients without all-cause mortality				
<5%	92	6 ^c		98
5%-10%	24 ^b	41	9 ^e	74
>10%	50	440	230	279
Total	121	91	239	451

Multivariate model adjusted for Framingham risk factors (recipient age and sex, systolic BP, antihypertensive treatment use, smoking status, diabetes mellitus, plasma total cholesterol and HDL cholesterol) with or without plasma C-terminal fibroblast growth factor 23 level. FGF23, fibroblast growth factor 23.

^aNet reclassification improvement was 0.07 (95% confidence interval [CI], 0.01 to 0.14; P<0.05).

^bNumbers of patients who were correctly reclassified by the model with FGF23.

^cNumbers of patients who were incorrectly reclassified by the model with FGF23.

^dNet reclassification improvement was 0.11 (95% CI, 0.05 to 0.18; P<0.001).

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Disclosures

None.

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