Regulation of Potassium Homeostasis

Biff F. Palmer

Abstract
Potassium is the most abundant cation in the intracellular fluid, and maintaining the proper distribution of potassium across the cell membrane is critical for normal cell function. Long-term maintenance of potassium homeostasis is achieved by alterations in renal excretion of potassium in response to variations in intake. Understanding the mechanism and regulatory influences governing the internal distribution and renal clearance of potassium under normal circumstances can provide a framework for approaching disorders of potassium commonly encountered in clinical practice. This paper reviews key aspects of the normal regulation of potassium metabolism and is designed to serve as a readily accessible review for the well-informed clinician as well as a resource for teaching trainees and medical students.


Introduction
Potassium plays a key role in maintaining cell function. Almost all cells possess an Na⁺-K⁺-ATPase, which pumps Na⁺ out of the cell and K⁺ into the cell and leads to a K⁺ gradient across the cell membrane (K⁺ᵢₘ > K⁺ᵢₒ) that is partially responsible for maintaining the potential difference across the membrane. This potential difference is critical to the function of cells, particularly in excitable tissues, such as nerve and muscle. The body has developed numerous mechanisms for defense of serum K⁺. These mechanisms serve to maintain a proper distribution of K⁺ within the body as well as regulate the total body K⁺ content.

Internal Balance of K⁺
The kidney is primarily responsible for maintaining total body K⁺ content by matching K⁺ intake with K⁺ excretion. Adjustments in renal K⁺ excretion occur over several hours; therefore, changes in extracellular K⁺ concentration are initially buffered by movement of K⁺ into or out of skeletal muscle. The regulation of K⁺ distribution between the intracellular and extracellular space is referred to as internal K⁺ balance. The most important factors regulating this movement under normal conditions are insulin and catecholamines (1).

After a meal, the postprandial release of insulin functions to not only regulate the serum glucose concentration but also shift dietary K⁺ into cells until the kidney excretes the K⁺ load re-establishing K⁺ homeostasis. These effects are mediated through insulin binding to cell surface receptors, which stimulates glucose uptake in insulin-responsive tissues through the insertion of the glucose transporter protein GLUT4 (2,3). An increase in the activity of the Na⁺-K⁺-ATPase mediates K⁺ uptake (Figure 1). In patients with the metabolic syndrome or CKD, insulin-mediated glucose uptake is impaired, but cellular K⁺ uptake remains normal (4,5), demonstrating differential regulation of insulin-mediated glucose and K⁺ uptake.

Catecholamines regulate internal K⁺ distribution, with α-adrenergic receptors impairing and β-adrenergic receptors promoting cellular entry of K⁺. β₂-Receptor-induced stimulation of K⁺ uptake is mediated by activation of the Na⁺-K⁺-ATPase pump. These effects play a role in regulating the cellular release of K⁺ during exercise (6).

Under normal circumstances, exercise is associated with movement of intracellular K⁺ into the interstitial space in skeletal muscle. Increases in interstitial K⁺ can be as high as 10–12 mM with severe exercise. Accumulation of K⁺ is a factor limiting the excitability and contractile force of muscle accounting for the development of fatigue (7,8). Additionally, increases in interstitial K⁺ play a role in eliciting rapid vaso-dilation, allowing for blood flow to increase in exercising muscle (9). During exercise, release of catecholamines through β₂ stimulation limits the rise in extracellular K⁺ concentration that otherwise occurs as a result of normal K⁺ release by contracting muscle. Although the mechanism is likely to be multifactorial, total body K⁺ depletion may blunt the accumulation of K⁺ into the interstitial space, limiting blood flow to skeletal muscle and accounting for the association of hypokalemia with rhabdomyolysis.

Changes in plasma tonicity and acid–base disorders also influence internal K⁺ balance. Hyperglycemia leads to water movement from the intracellular to extracellular compartment. This water movement favors K⁺ efflux from the cell through the process of solvent drag. In addition, cell shrinkage causes intracellular K⁺ concentration to increase, creating a more favorable concentration gradient for K⁺ efflux. Mineral acidosis, but not organic acidosis, can be a cause of cell shift in K⁺. As recently reviewed, the general effect of acidemia to cause K⁺ loss from cells is not because of a direct K⁺-H⁺ exchange, but, rather, is because of an apparent coupling resulting from effects of acidosis on transporters that normally regulate cell pH in skeletal muscle (10) (Figure 2).
Intracellular K⁺ serves as a reservoir to limit the fall in extracellular K⁺ concentrations occurring under pathologic conditions where there is loss of K⁺ from the body. The efficiency of this effect was shown by military recruits undergoing training in the summer (11). These subjects were able to maintain a near-normal serum K⁺ under conditions of K⁺ loss of cellular K⁺ serves as a reservoir to limit the fall in extracellular K⁺ concentration that occur under conditions of systemic acidosis. Inorganic anion and H⁺ through the monocarboxylate transporter (MCT; MCT1 and MCT4). Accumulation of the acid results in a larger fall in intracellular pH, thereby stimulating inward movement of Cl⁻ by Cl⁻-HCO₃⁻ exchange, further enhancing K⁺ efflux by K⁺-Cl⁻ cotransport. (B) Loss of K⁺ from the extracellular space of skeletal muscle.

In rats deprived of K⁺ for 10 days, the plasma K⁺ concentration decreased from 4.2 to 2.9 mmol/L. Insulin-mediated K⁺ disappearance declined by more than 90% compared with control values. This decrease in K⁺ uptake was accompanied by a >50% reduction in both the activity and expression of muscle Na⁺-K⁺-ATPase, suggesting that decreased pump activity might account for the decrease in insulin effect. This decrease in muscle K⁺ uptake, under conditions of K⁺ depletion, may limit excessive falls in extracellular K⁺ concentration that occur under conditions of insulin stimulation. Concurrently, reductions in pump expression and activity facilitate the ability of skeletal muscle to buffer declines in extracellular K⁺ concentrations by donating some component of its intracellular stores.

There are differences between skeletal and cardiac muscle in the response to chronic K⁺ depletion. Although skeletal muscle readily relinquishes K⁺ to minimize the drop in plasma K⁺ concentration, cardiac tissue K⁺ content remains relatively well preserved. In contrast to the decline in activity and expression of skeletal muscle Na⁺-K⁺-ATPase, cardiac Na⁺-K⁺-ATPase pool size

![Figure 1.](image1.png) The cell model illustrates β₂-adrenergic and insulin-mediated regulatory pathways for K⁺ uptake. β₂-Adrenergic and insulin both lead to K⁺ uptake by stimulating the activity of the Na⁺-K⁺-ATPase pump primarily in skeletal muscle, but they do so through different signaling pathways. β₂-Adrenergic stimulation leads to increased pump activity through a cAMP- and protein kinase A (PKA)-dependent pathway. Insulin binding to its receptor leads to phosphorylation of the insulin receptor substrate protein (IRS-1), which, in turn, binds to phosphatidylinositol 3-kinase (PI3-K). The IRS-1–PI3-K interaction leads to activation of 3-phosphoinositide-dependent protein kinase-1 (PDK1). The stimulatory effect of insulin on glucose uptake and K⁺ uptake diverge at this point. An Akt-dependent pathway is responsible for membrane insertion of the glucose transporter GLUT4, whereas activation of atypical protein kinase C (aPKC) leads to membrane insertion of the Na⁺-K⁺-ATPase pump (reviewed in ref. 3).

![Figure 2.](image2.png) The effect of metabolic acidosis on internal K⁺ balance in skeletal muscle. (A) In metabolic acidosis caused by inorganic anions (mineral acidosis), the decrease in extracellular pH will decrease the rate of Na⁺-H⁺ exchange (NHE1) and inhibit the inward rate of Na⁺-3HCO₃⁻ cotransport (NBCe1 and NBCe2). The resultant fall in intracellular Na⁺ will reduce Na⁺-K⁺-ATPase activity, causing a net loss of cellular K⁺. In addition, the fall in extracellular HCO₃⁻ concentration will increase inward movement of Cl⁻ by Cl⁻-HCO₃⁻ exchange, further enhancing K⁺ efflux by K⁺-Cl⁻ cotransport. (B) Loss of K⁺ from the cell is much smaller in magnitude in metabolic acidosis caused by an organic acidosis. In this setting, there is a strong inward flux of the organic anion and H⁺ through the monocarboxylate transporter (MCT; MCT1 and MCT4). Accumulation of the acid results in a larger fall in intracellular pH, thereby stimulating inward movement of Na⁺ by way of Na⁺-H⁺ exchange and Na⁺-3HCO₃⁻ cotransport. Accumulation of intracellular Na⁺ maintains Na⁺-K⁺-ATPase activity, thereby minimizing any change in extracellular K⁺ concentration.

![Image](image3.png)
increases in K⁺-deficient animals. This difference explains the greater total K⁺ clearance capacity after the acute administration of intravenous KCl to rats fed a K⁺-free diet for 2 weeks compared with K⁺-replete controls (13,14). Cardiac muscle accumulates a considerable amount of K⁺ in the setting of an acute load. When expressed on a weight basis, the cardiac capacity for K⁺ uptake is comparable with that of skeletal muscle under conditions of K⁺ depletion and may actually exceed skeletal muscle under control conditions.

**Renal Potassium Handling**

Potassium is freely filtered by the glomerulus. The bulk of filtered K⁺ is reabsorbed in the proximal tubule and loop of Henle, such that less than 10% of the filtered load reaches the distal nephron. In the proximal tubule, K⁺ absorption is primarily passive and proportional to Na⁺ and water (Figure 3). K⁺ reabsorption in the thick ascending limb of Henle occurs through both transcellular and paracellular pathways. The transcellular component is mediated by K⁺ transport on the apical membrane Na⁺-K⁺-2Cl⁻ cotransporter (Figure 4). K⁺ secretion begins in the early distal convoluted tubule and progressively increases along the distal nephron into the cortical collecting duct (Figure 5). Most urinary K⁺ can be accounted for by electrogenic K⁺ secretion mediated by principal cells in the initial collecting duct and the cortical collecting duct (Figure 6). An electroneutral K⁺ and Cl⁻ cotransport mechanism is also present on the apical surface of the distal nephron (15). Under conditions of K⁺ depletion, reabsorption of K⁺ occurs in the collecting duct. This process is mediated by upregulation in the apically located H⁺-K⁺-ATPase on α-intercalated cells (16) (Figure 7).

Under most homeostatic conditions, K⁺ delivery to the distal nephron remains small and is fairly constant. By contrast, the rate of K⁺ secretion by the distal nephron varies and is regulated according to physiologic needs. The cellular determinants of K⁺ secretion in the principal cell include the intracellular K⁺ concentration, the luminal K⁺ concentration, the potential (voltage) difference across the luminal membrane, and the permeability of the luminal membrane for K⁺. Conditions that increase cellular K⁺ concentration, decrease luminal K⁺ concentration, or render the lumen more electronegative will increase the rate of K⁺ secretion. Conditions that increase the permeability of the luminal membrane for K⁺ will increase the rate of K⁺ secretion. Two principal determinants of K⁺ secretion are mineralocorticoid activity and distal delivery of Na⁺ and water.

Aldosterone is the major mineralocorticoid in humans and affects several of the cellular determinants discussed above, leading to stimulation of K⁺ secretion. First, aldosterone increases intracellular K⁺ concentration by stimulating the activity of the Na⁺-K⁺-ATPase in the basolateral membrane. Second, aldosterone stimulates Na⁺ reabsorption across the luminal membrane, which increases the electronegativity of the lumen, thereby increasing the electrical gradient favoring K⁺ secretion. Lastly, aldosterone has a direct effect on the luminal membrane to increase K⁺ permeability (17).

---

**Figure 3.** | A cell model for K⁺ transport in the proximal tubule. K⁺ reabsorption in the proximal tubule primarily occurs through the paracellular pathway. Active Na⁺ reabsorption drives net fluid reabsorption across the proximal tubule, which in turn, drives K⁺ reabsorption through a solvent drag mechanism. As fluid flows down the proximal tubule, the luminal voltage shifts from slightly negative to slightly positive. The shift in transepithelial voltage provides an additional driving force favoring K⁺ diffusion through the low-resistance paracellular pathway. Experimental studies suggest that there may be a small component of transcellular K⁺ transport; however, the significance of this pathway is not known. K⁺ uptake through the Na⁺-K⁺-ATPase pump can exit the basolateral membrane through a conductive pathway or coupled to Cl⁻. An apically located K⁺ channel functions to stabilize the cell negative potential, particularly in the setting of Na⁺-coupled cotransport of glucose and amino acids, which has a depolarizing effect on cell voltage.

**Figure 4.** | A cell model for K⁺ transport in the thick ascending limb of Henle. K⁺ reabsorption occurs by both paracellular and transcellular mechanisms. The basolateral Na⁺-K⁺-ATPase pump maintains intracellular Na⁺ low, thus providing a favorable gradient to drive the apically located Na⁺-K⁺-2Cl⁻ cotransporter (an example of secondary active transport). The apically located renal outer medullary K⁺ (ROMK) channel provides a pathway for K⁺ to recycle from cell to lumen, and ensures an adequate supply of K⁺ to sustain Na⁺-K⁺-2Cl⁻ cotransport. This movement through ROMK creates a lumen-positive voltage, providing a driving force for passive K⁺ reabsorption through the paracellular pathway. Some of the K⁺ entering the cell through the cotransporter exits the cell across the basolateral membrane, accounting for transcellular K⁺ reabsorption. K⁺ can exit the cell through a conductive pathway or in cotransport with Cl⁻. CIC-Kb is the primary pathway for Cl⁻ efflux across the basolateral membrane.
mediated K1tisone, the latter of which has no affinity for the receptor. Electrogenic-receptor free to only bind aldosterone by metabolizing cortisol to cor-
dehydrogenase II. This enzyme maintains the mineralocorticoid
t h er a t eo fd i s t a ld e l i v e yo fN a
absorbable anions, such as sulfate, phosphate, or bicarbonate.

Figure 5. | A cell model for K+ transport in the distal convoluted
tubule (DCT). In the early DCT, luminal Na+ uptake is mediated by the
apically located thiazide-sensitive Na+-Cl- cotransporter. The
transporter is energized by the basolateral Na+-K+-ATPase, which
maintains intracellular Na+ concentration low, thus providing a fa-
vorable gradient for Na+ entry into the cell through secondary active
transport. The cotransporter is abundantly expressed in the DCT1
but progressively declines along the DCT2. ROMK is expressed
throughout the DCT and into the cortical collecting duct. Expression
of the epithelial Na+ channel (ENaC), which mediates amiloride-
sensitive Na+ absorption, begins in the DCT2 and is robustly ex-
pressed throughout the downstream connecting tubule and cortical
collecting duct. The DCT2 is the beginning of the aldosterone-
sensitive distal nephron (ASDN) as identified by the presence of both
the mineralocorticoid receptor and the enzyme 11β-hydroxysteroid
dehydrogenase II. This enzyme maintains the mineralocorticoid
receptor free to only bind aldosterone by metabolizing cortisol to cor-
tisone, the latter of which has no affinity for the receptor. Electrogenic-
mediated K+ transport begins in the DCT2 with the combined presence
of ROMK, ENaC, and aldosterone sensitivity. Electroneutral K+-Cl-
cotransporter in the DCT and collecting duct. Conditions
that cause a low luminal Cl- concentration increase K+ secretion
through this mechanism, which occurs with delivery of poorly re-
absorbable anions, such as sulfate, phosphate, or bicarbonate.

A second principal determinant affecting K+ secretion is the rate of distal delivery of Na+ and water. Increased
distal delivery of Na+ stimulates distal Na+ absorption, which will make the luminal potential more negative
and, thus, increase K+ secretion. Increased flow rates also increase K+ secretion. When K+ is secreted in the
collecting duct, the luminal K+ concentration rises, which decreases the diffusion gradient and slows additional K+
secretion. At higher luminal flow rates, the same amount of K+ secretion will be diluted by the larger volume such
that the rise in luminal K+ concentration will be less. Thus,
increases in the distal delivery of Na+ and water stimulate
K+ secretion by lowering luminal K+ concentration and
making the luminal potential more negative.

Two populations of K+ channels have been identified in the
cells of the cortical collecting duct. The renal outer medullary
K+ (ROMK) channel is considered to be the major K+-secretory
pathway. This channel is characterized by having low con-
ductance and a high probability of being open under phys-
ologic conditions. The maxi-K+ channel (also known as the
large-conductance K+ [BK] channel) is characterized by a
large single channel conductance and quiescence in the basal
state and activation under conditions of increased flow (18).
In addition to increased delivery of Na+ and dilution of
luminal K+ concentration, recruitment of maxi-K+ channels
contributes to flow-dependent increased K+ secretion. Renal
K+ channels are subjects of extensive reviews (19–21).

Figure 6. | The cell that is responsible for K+ secretion in the initial
collecting duct and the cortical collecting duct is the principal cell.
This cell possesses a basolateral Na+-K+-ATPase that is responsible
for the active transport of K+ from the blood into the cell. The resultant
high cell K+ concentration provides a favorable diffusion gradient for
movement of K+ from the cell into the lumen. In addition to estab-
lishing a high intracellular K+ concentration, activity of this pump
lowers intracellular Na+ concentration, thus maintaining a favorable
diffusion gradient for movement of Na+ from the lumen into the cell.
Both the movements of Na+ and K+ across the apical membrane
occur through well defined Na+ and K+ channels.

Figure 7. | Reabsorption of HCO3 in the distal nephron is mediated
by apical H+ secretion by the α-intercalated cell. Two transporters
secrete H+, a vacuolar H+-ATPase and an H+-K+-ATPase. The H+-K+
-ATPase uses the energy derived from ATP hydrolysis to secrete H+
into the lumen and reabsorb K+ in an electroneutral fashion. The
activity of the H+-K+-ATPase increases in K+ depletion and, thus,
provides a mechanism by which K+ depletion enhances both col-
lecting duct H+ secretion and K+ absorption.
The effect of increased tubular flow to activate maxi-K\(^+\) channels may be mediated by changes in intracellular Ca\(^{2+}\) concentration (22). The channel is Ca\(^{2+}\)-activated, and an acute increase in flow increases intracellular Ca\(^{2+}\) concentrations in the principal cell. It has been suggested that the central cilium (a structure present in principal cells) may facilitate transduction of signals of increased flow to increased intracellular Ca\(^{2+}\) concentration. In cultured cells, bending of primary cilia results in a transient increase in intracellular Ca\(^{2+}\), an effect blocked by antibodies to polycystin 2 (23). Although present in nearly all segments of the nephron, the maxi-K channel has been identified as the mediator of flow-induced K\(^+\) secretion in the distal nephron and cortical collecting duct (24).

Development of hypokalemia in type II Bartter syndrome illustrates the importance of maxi-K channels in renal K\(^+\) excretion (25). Patients with type II Bartter syndrome have a loss-of-function mutation in ROMK manifesting with clinical features of the disease in the perinatal period. ROMK provides the pathway for recycling of K\(^+\) across the apical membrane in the thick ascending limb of Henle. This recycling generates a lumen-positive potential that drives the paracellular reabsorption of Ca\(^{2+}\) and Mg\(^{2+}\) and provides luminal K\(^+\) to the Na\(^+-K^+-2Cl^-\) co-transporter (Figure 4).

Mutations in ROMK decrease NaCl and fluid reabsorption in the thick limb, mimicking a loop diuretic effect, which causes volume depletion. Despite the increase in distal Na\(^+\) delivery, K\(^+\)-wasting is not consistently observed, because ROMK is also the major K\(^+\)-secretory pathway for regulated K\(^+\) excretion in the collecting duct. In fact, in the perinatal period, infants with this form of Bartter syndrome often exhibit a transient hyperkalemia consistent with loss of function of ROMK in the collecting duct. Over time, these patients develop hypokalemia as a result of increased flow-mediated K\(^+\) secretion through maxi-K\(^+\) channels. Studies in an ROMK-deficient mouse model of type II Bartter syndrome are consistent with this mechanism (26). The transient hyperkalemia observed in the perinatal period is likely related to the fact that ROMK channels are functionally expressed earlier than maxi-K\(^+\) channels during the course of development.

In this regard, growing infants and children are in a state of positive K\(^+\) balance, which correlates with growth and increasing cell number. Early in development, there is a limited capacity of the distal nephron to secrete K\(^+\) because of a paucity of both apically located ROMK and maxi-K\(^+\) channels. The increase in K\(^+\)-secretory capacity with maturation is initially a result of increased expression of ROMK. Several weeks later, maxi-K\(^+\) channel expression develops, allowing for flow-mediated K\(^+\) secretion to occur (reviewed in ref. 27). The limitation in distal K\(^+\) secretion is channel-specific, because the electrochemical gradient favoring K\(^+\)-secretion, as determined by activity of the Na\(^+-K^+-\)ATPase and Na\(^+\) reabsorption, is not limiting. Additionally, increased flow rates are accompanied by appropriate increases in Na\(^+\) reabsorption and intracellular Ca\(^{2+}\) concentrations in the distal nephron, despite the absence of stimulatory effect on K\(^+\) secretion (28). Activity of the H\(^+\)-K\(^+-\)ATPase, which couples K\(^+\) reabsorption to H\(^+\) secretion in intercalated cells, is similar in newborns and adults. K\(^+\) reabsorption through this pump, combined with decreased expression of K\(^+\)-secretory channels, helps maintain a state of positive K\(^+\) balance during somatic growth after birth. These features of distal K\(^+\) handling by the developing kidney are a likely explanation for the high incidence of nonoliguric hyperkalemia in preterm infants (29).

Another physiologic state characterized by a period of positive K\(^+\) balance is pregnancy, where approximately 300 mEq K\(^+\) is retained (30). High circulating levels of progesterone may play a role in this adaptation through stimulatory effects on K\(^+\) and H\(^+\) transport by the H\(^+\)-K\(^+\) \(\alpha_2\)-ATPase isoform in the distal nephron (31).

In addition to stimulating maxi-K\(^+\) channels, increased tubular flow has been shown to stimulate Na\(^+\) absorption through the epithelial Na\(^+\) channel (ENaC) in the collecting duct. This increase in absorption not only is because of increased delivery of Na\(^+\), but also seems to be the result of mechanosensitive properties intrinsic to the channel. Increased flow creates a shear stress that activates ENaCs by increasing channel open probability (32,33).

It has been hypothesized that biomechanical regulation of renal tubular Na\(^+\) and K\(^+\) transport in the distal nephron may have evolved as a response to defend against sudden increases in extracellular K\(^+\) concentration that occur in response to ingestion of K\(^+\)-rich diets typical of early vertebrates (22). According to this hypothesis, an increase in GFR after a protein-rich meal would lead to an increase in distal flow activating the ENaC, increasing intracellular Ca\(^{2+}\) concentration, and activating maxi-K\(^+\) channels. These events would enhance K\(^+\) secretion, thus providing a buffer to guard against development of hyperkalemia.

In patients with CKD, loss of nephron mass is counterbalanced by an adaptive increase in the secretory rate of K\(^+\) in remaining nephrons such that K\(^+\) homeostasis is generally well maintained until the GFR falls below 15–20 ml/min (34). The nature of the adaptive process is thought to be similar to the adaptive process that occurs in response to high dietary K\(^+\) intake in normal subjects (35). Chronic K\(^+\) loading in animals augments the secretory capacity of the distal nephron, and, therefore, renal K\(^+\) excretion is significantly increased for any given plasma K\(^+\) level. Increased K\(^+\) secretion under these conditions occurs in association with structural changes characterized by cellular hypertrophy, increased mitochondrial density, and proliferation of the basolateral membrane in cells in the distal nephron and principal cells of the collecting duct. Increased serum K\(^+\) and mineralocorticoids independently initiate the amplification process, which is accompanied by an increase in Na\(^+-K^+-\)ATPase activity.

**Aldosterone Paradox**

Under conditions of volume depletion, activation of the renin-angiotensin system leads to increased aldosterone release. The increase in circulating aldosterone stimulates renal Na\(^+\) retention, contributing to the restoration of extracellular fluid volume, but occurs without a demonstrable effect on renal K\(^+\) secretion. Under condition of hyperkalemia, aldosterone release is mediated by a direct effect of K\(^+\) on cells in the zona glomerulosa. The subsequent increase in circulating aldosterone stimulates renal K\(^+\) secretion, restoring the serum K\(^+\) concentration to normal, but does so without concomitant renal Na\(^+\) retention.
The ability of aldosterone to signal the kidney to stimulate salt retention without K\(^+\) secretion in volume depletion and stimulate K\(^+\) secretion without salt retention in hyperkalemia has been referred to as the aldosterone paradox (36). In part, this ability can be explained by the reciprocal relationship between urinary flow rates and distal Na\(^+\) delivery with circulating aldosterone levels. Under conditions of volume depletion, proximal salt and water absorption increase, resulting in decreased distal delivery of Na\(^+\) and water. Although aldosterone levels are increased, renal K\(^+\) excretion remains fairly constant, because the stimulatory effect of increased aldosterone is counterbalanced by the decreased delivery of filtrate to the distal nephron. Under condition of an expanded extracellular fluid volume, distal delivery of filtrate is increased as a result of decreased proximal tubular fluid reabsorption. Once again, renal K\(^+\) excretion remains relatively constant in this setting, because circulating aldosterone levels are suppressed. It is only under pathophysiologic conditions that increased distal Na\(^+\) and water delivery are coupled to increased aldosterone levels. Renal K\(^+\) wasting will occur in this setting (37) (Figure 8).

Renal K\(^+\) secretion also remains stable during changes in flow rate resulting from variations in circulating vasopressin. In this regard, vasopressin has a stimulatory effect on renal K\(^+\) secretion (38,39). This kaliuretic property may serve to oppose a tendency to K\(^+\) retention under conditions of antidiuresis when a low-flow rate-dependent fall in distal tubular K\(^+\) secretion might otherwise occur. In contrast, suppressed endogenous vasopressin leads to decreased activity of the distal K\(^+\)-secretory mechanism, thus limiting excessive K losses under conditions of full hydration and water diuresis.

Although the inverse relationship between aldosterone levels and distal delivery of salt and water serves to keep renal K\(^+\) excretion independent of volume status, recent reviews have suggested a more complex mechanism centered on the with no lysine [K]\(^4\) (WNK4) protein kinase in the distal nephron (40,41). WNK4 is one of four members of a family of serine-threonine kinases each encoded by a different gene and characterized by the atypical placement of the catalytic lysine residue that is present in most other protein kinases. Inactivating mutations in WNK4 lead to development of pseudohypoaldosteronism type II (PHAII; Gordon syndrome). This disorder is inherited in an autosomal dominant fashion and is characterized by hypertension and hyperkalemia (42). Circulating aldosterone levels are low, despite the presence of hyperkalemia. Thiazide diuretics are particularly effective in treating both the hypertension and hyperkalemia (43).

Wild-type WNK4 acts to reduce surface expression of the thiazide-sensitive Na\(^+\)-Cl\(^-\) cotransporter and also stimulates clathrin-dependent endocytosis of ROMK in the collecting duct (44,45). The inactivating mutation of WNK4 responsible for PHAII leads to increased cotransporter activity and further stimulates endocytosis of ROMK. The net effect is increased NaCl reabsorption combined with decreased K\(^+\) secretion. Mutated WNK4 also enhances paracellular Cl\(^-\) permeability caused by increased phosphorylation of claudins, which are tight junction proteins involved in regulating paracellular ion transport (46). In addition to increasing Na\(^+\) retention, this change in permeability further impairs K\(^+\) secretion, because the lumen-negative voltage, which normally serves as a driving force for K\(^+\) secretion, is dissipated.

Because development of hypertension and hyperkalemia resulting from the PHAII-mutated WNK4 protein can be viewed as an exaggerated response to a reduction in extracellular fluid volume (salt retention without increased K\(^+\) secretion), it has been proposed that wild-type WNK4 may act as a molecular switch determining balance between renal NaCl reabsorption and K\(^+\) secretion (45,47). Under conditions of volume depletion, the switch would be altered in a manner reminiscent of the PHAII mutant such that NaCl reabsorption is increased, but K\(^+\) secretion is further

![Figure 8](image)

**Figure 8.** Under normal circumstances, delivery of Na\(^+\) to the distal nephron is inversely associated with serum aldosterone levels. For this reason, renal K\(^+\) excretion is kept independent of changes in extracellular fluid volume. Hypokalemia caused by renal K\(^+\) wasting can be explained by pathophysiologic changes that lead to coupling of increased distal Na\(^+\) delivery and aldosterone or aldosterone-like effects. When approaching the hypokalemia caused by renal K\(^+\) wasting, one must determine whether the primary disorder is an increase in mineralocorticoid activity or an increase in distal Na\(^+\) delivery. EABV, effective arterial blood volume.
inhibited. However, when increased serum K⁺ concentration occurs in the absence of volume depletion, WNK4 alterations result in maximal renal K⁺ secretion without Na⁺ retention.

Angiotensin II (AII) has emerged as an important modulator of this switch. Under conditions of volume depletion, AII and aldosterone levels are increased (Figure 9). In addition to effects leading to enhanced NaCl reabsorption in the proximal tubule, AII activates the Na⁺-Cl⁻ cotransporter in a WNK4-dependent manner, and it is primarily located in the initial part of the distal convoluted tubule (DCT; DCT1) (48,49). AII also activates ENaC, which is found in the aldosterone-sensitive distal nephron (ASDN) comprised of the second segment of the DCT (DCT2), the connecting tubule, and the collecting duct (50). The activation of ENaC by AII is additive to that of aldosterone (51). In this manner, AII and aldosterone act in concert to stimulate Na⁺ retention. At the same time, AII inhibits ROMK by both WNK4-dependent and -independent mechanisms (52,53). This inhibitory effect on ROMK along with decreased Na⁺ delivery to the collecting duct brought about by AII stimulation of Na⁺ reabsorption in the proximal nephron, and DCT1 allows for simultaneous Na⁺ conservation without K⁺ wasting.

Hyperkalemia, or an increase in dietary K⁺ intake, can increase renal K⁺ secretion independent of change in mineralocorticoid activity and without causing volume retention. This effect was shown in Wistar rats fed a diet very low in NaCl and K⁺ for several days and given a pharmacologic dose of deoxycorticosterone to ensure a constant serum K⁺ concentration (48). AII also activates ENaC and aldosterone. As discussed in the text, with no lysine [K] 4 (WNK4) proteins are integrally involved in the signals by which the paradox is brought about. It should be emphasized that the WNK proteins are part of a complex signaling network still being fully elucidated. The interested reader is referred to several recent reviews and advancements on this subject (48,51,91–93).

The aldosterone paradox refers to the ability of the kidney to stimulate NaCl retention with minimal K⁺ secretion under conditions of volume depletion and maximize K⁺ secretion without Na⁺ retention in hyperkalemia. With volume depletion (left panel), increased circulating angiotensin II (AII) levels stimulate the Na⁺-Cl⁻ cotransporter in the early DCT. In the ASDN, AII along with aldosterone stimulate the ENaC. In this latter segment, AII exerts an inhibitory effect on ROMK, thereby providing a mechanism to maximally conserve salt and minimize renal K⁺ secretion. When hyperkalemia or increased dietary K⁺ intake occurs with normovolemia (right panel), low circulating levels of AII or direct effects of K⁺ lead to inhibition of Na⁺-Cl⁻ cotransporter activity along with increased activity of ROMK. As a result, Na⁺ delivery to the ENaC is optimized for the coupled electrogenic secretion of K⁺ through ROMK. As discussed in the text, with no lysine [K] 4 (WNK4) proteins are integrally involved in the signals by which the paradox is brought about. It should be emphasized that the WNK proteins are part of a complex signaling network still being fully elucidated. The interested reader is referred to several recent reviews and advancements on this subject (48,51,91–93).
In this regard, increased dietary K⁺ intake leads to an inhibitory effect on Na⁺ transport in this segment and does so through effects on WNK1, another member of the WNK family of kinases (60,61). WNK1 is ubiquitously expressed throughout the body in multiple spliced forms. By contrast, a shorter WNK1 transcript lacking the amino terminal 1–437 amino acids of the long transcript is highly expressed in the kidney but not other tissues, and it is referred to as kidney-specific WNK1 (KS-WNK1). KS-WNK1 is restricted to the DCT and part of the connecting duct and functions as a physiologic antagonist to the actions of long WNK1. Changes in the ratio of KS-WNK1 and long WNK1 in response to dietary K⁺ contribute to the physiologic regulation of renal K⁺ excretion (62–65).

Under normal circumstances, long WNK1 prevents the ability of WNK4 to inhibit activity of the Na⁺–Cl⁻ cotransporter in the DCT. Thus, increased activity of long WNK1 leads to a net increase in NaCl reabsorption. Dietary K⁺ loading increases the abundance of KS-WNK1. Increased KS-WNK1 antagonizes the inhibitory effect of long WNK1 on WNK4. The net effect is inhibition of Na⁺–Cl⁻ cotransport in the DCT and increased Na⁺ delivery to more distal parts of the tubule. In addition, increased KS-WNK1 antagonizes the effect of long WNK1 to stimulate endocytosis of ROMK. Furthermore, KS-WNK1 exerts a stimulatory effect on the ENaC. Thus, increases in KS-WNK1 in response to dietary K⁺ loading facilitate K⁺ secretion through the combined effects of increased Na⁺ delivery through downregulation of Na⁺–Cl⁻ cotransport in the DCT, increased electrogenic Na⁺ reabsorption through the ENaC, and greater abundance of ROMK.

Increased aldosterone levels in response to a high K⁺ diet lead to effects that complement the effects of KS-WNK1 (66,67). The serum- and glucocorticoid-dependent protein kinase (SGK1) is an immediate transcriptional target of aldosterone binding to the mineralocorticoid receptor. Activation of SGK1 leads to phosphorylation of WNK4, resulting in a loss of the ability of WNK4 to inhibit ROMK and the phosphorylation of ubiquitin protein ligase Nedd4. The phosphorylation of ubiquitin protein ligase Nedd4 results in increased ENaC expression and activity by causing ENaC (66,68). Aldosterone-induced activation of SGK1 also antagonizes the effect of long WNK1 to stimulate endocytosis of ROMK. Furthermore, KS-WNK1 exerts a stimulatory effect on the ENaC by causing ENaC (66,67). The serum- and glucocorticoid-dependent protein kinase functions as a physiologic antagonist to the actions of long WNK1.

Role in Hypertension

Changes in KS-WNK1 and long WNK1 that occur in response to dietary K⁺ intake affect renal Na⁺ handling in a way that may be of importance in the observed relationship between dietary K⁺ intake and hypertension. Epidemiologic studies established that K⁺ intake is inversely related to the prevalence of hypertension (70). In addition, K⁺ supplements and avoidance of hypokalemia lowers BP in hypertensive subjects. By contrast, BP increases in hypertensive subjects placed on a low K⁺ diet. This increase in BP is associated with increased renal Na⁺ reabsorption (71).

K⁺ deficiency increases the ratio of long WNK1 to KS-WNK1. Long WNK1 is associated with increased retrieval of ROMK, thus providing an appropriate response to limit K⁺ secretion. However, long WNK1 also leads to a stimulatory effect on ENaC activity as well as releasing the inhibitory effect of WNK4 on Na⁺ reabsorption mediated by the NaCl cotransporter in the DCT (72,73). These effects suggest that reductions in K⁺ secretion under conditions of K⁺ deficiency will occur at the expense of increased Na⁺ retention.

Renal conservation of K⁺ and Na⁺ under conditions of K⁺ deficiency may be considered an evolutionary adaptation, because dietary K⁺ and Na⁺ deficiency likely occurred together for early humans (74). However, such an effect is potentially deleterious in our present setting, because evolution has seen a large increase in the ratio of dietary intake of Na⁺ versus K⁺. The effects of an increased ratio of WNK1 to KS-WNK1 in the kidney under conditions of modern day high Na⁺/low K⁺ diet could be central to the pathogenesis of salt-sensitive hypertension (75).

Enteric Sensor of K⁺

There is evidence to support the existence of enteric solute sensors capable of responding to dietary Na⁺, K⁺, and phosphate that signal the kidney to rapidly alter ion excretion or reabsorption (76–78). In experimental animals, and using protocols to maintain identical plasma K⁺ concentration, the kaliuretic response to a K⁺ load is greater when given as a meal compared with an intravenous infusion (79). These studies suggest that dietary K⁺ intake through a splanchnic sensing mechanism can signal increases in renal K⁺ excretion independent of changes in plasma K⁺ concentration or aldosterone (reviewed in ref. 80).

Although the precise signaling mechanism is not known, recent studies suggest that the renal response may be because of rapid and nearly complete dephosphorylation of the Na⁺–Cl⁻ cotransporter in the DCT, causing decreased activity of the transporter and, thus, enhancing delivery of Na⁺ to the ASDN (81,82). In these studies, gastric delivery of K⁺ led to dephosphorylation of the cotransporter within minutes independent of aldosterone and based on in vitro studies, independent of changes in extracellular K⁺ concentration. The temporally associated increase in renal K⁺ excretion results from a more favorable electrochemical driving force caused by the downstream shift in Na⁺ reabsorption from the DCT to the ENaC in the ASDN as well as increased maxi-K⁺ channel K⁺ secretion brought on by increased flow. This rapid natriuretic response to increases in dietary K⁺ intake is consistent with the BP-lowering effect of K⁺-rich diets discussed earlier.

Circadian Rhythm of K⁺ Secretion

During a 24-hour period, urinary K⁺ excretion varies in response to changes in activity and fluctuations in K⁺ intake caused by the spacing of meals. However, even when K⁺ intake and activity are evenly spread over a 24-hour period, there remains a circadian rhythm whereby K⁺ excretion is lower at night and in the early morning hours and then increases in the afternoon (83–86). This circadian pattern results from changes in intratubular K⁺ concentration in the collecting duct as opposed to variations in urine flow rate (87).

In the mouse distal nephron, a circadian rhythm exists for gene transcripts that encode proteins involving K⁺
secretion (88). Gene expression of ROMK is greater during periods of activity, whereas expression of the H⁺-K⁺-AT-Pase is higher during rest, which corresponds to periods when renal K⁺ excretion is greater and less, respectively (89). Changes in plasma aldosterone levels may play a contributory role, because circadian rhythm of glucocorticoid synthesis and secretion has been described in the adrenal gland. In addition, expression of clock genes within cells of the distal nephron suggests that a pacemaker function regulating K⁺ transport may be an intrinsic component of the kidney that is capable of operating independent of outside influence.

The clinical significance of this rhythmicity in K⁺ and other electrolyte secretions is not known. Evidence suggests that dysregulation of circadian rhythms may contribute to a lack of nocturnal decline in BP, with eventual development of sustained hypertension as well as accelerated CKD and cardiovascular disease (85,90).

Disclosures
None.

References
42. Mayan H, Veere KD, Mouallem M, Tzadok-Witkon M, Pouwer N, Farfel Z: Pseudohypopaldosteronism type II: Marked sensitivity to


Published online ahead of print. Publication date available at www.cjasn.org.